



## Review article

# Revolutionizing of bioactive natural products in prostate cancer research and care: Promising discoveries and future directions

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## ABSTRACT

Globally, prostate cancer (PCa) is one of the most common cancers to strike men. Diet and lifestyle appear to have a significant impact on PCa biology and carcinogenesis. PCa is the major reason of death by cancer in men. Anti-PCa qualities like growth of tumor inhibition, induction of cell death, and angiogenesis and metastasis inhibition have all been studied in depth. Phytochemicals have been demonstrated to target androgen receptor (AR) signaling as well as PCa stem cells in a selection of investigations. Marine compounds have shown potential in the treatment of PCa. It is discussed in this article, some of the most promising bioactive natural and marine compounds for PCa prevention and treatment, as well as their specific methods of action. An emphasis on specific medicine is one of the future directions in the revolutionization of bioactive natural ingredients for PCa research and therapy. Advances in nanotechnology can enhance the bioavailability and specificity of bioactive substances for cancer cells, maximizing their therapeutic potential and enhancing patient treatment. Bioactive natural compounds represent an innovative field in the study and treatment of PCa. Promising results point to their potential to block cancer pathways and improve on already effective therapeutic approaches. As we advance, modified medicine, nanotechnology, and genomics methods will be fundamental in maximizing the efficacy of these natural substances and ultimately changing the treatment of PCa. But in order to close the gap between exciting findings and therapeutic application, more study, clinical trials, and effective activities are essential.

## 1. Introduction

Prostate cancer (PCa) is the most frequent cancer among males worldwide, with a high prevalence in Western nations.<sup>1</sup> In nearly ninety

percent of patients, PCa is still only locally progressed or organ-confined when they are diagnosed, making prostatectomy or local radiation treatment options,<sup>2</sup> and androgen-deprivation therapy, which tries to reduce hormone output and/or activity, is the most effective treatment at

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this point, when tumor growth is dependent on androgens. This treatment is pharmacological castration based, which is accomplished by using GnRH agonists in conjunction with antiandrogens or alone<sup>3,4</sup>; recent times, 2 large clinical trials, STAMPEDE and CHAARTED, have proven the advantages of combining hormonal therapy with chemotherapy.<sup>5,6</sup> Even after a positive first reaction, the vast proportion of patients relapse two to three years later, with the tumor expanding to the point that castration is no longer effective.<sup>7</sup> Better treatment options are necessary because chemotherapy and immunotherapy, & novel drugs like abiraterone and enzalutamide, only provide a few months of progress. Patients who are castration resistant have a better probability of survival if they are not castrated.<sup>4,8</sup> Bone metastases, which account for Eighty percent of developed PCa and are commonly dealt with chemotherapy and radiation, are linked to a high rate of morbidity, a poor quality of life, and a variety of skeletal complications.<sup>6,9</sup> Natural chemicals have attracted a lot of attention in recent years as a result of their unique anti-cancer potential. Nutraceuticals have an anti-metastatic, anti-angiogenic, pro-death, and growth-suppressing effect in cell lines of PCa that xenograft, but normal prostate epithelial cells are spared.<sup>10</sup> These medications' anti-PCa effects are attributable to a variety of mechanisms, including androgen receptor (AR) axis suppression as well as cancer stemness targeting.<sup>11–13</sup> As a source of natural medicine ingredients, marine sources have played an important part in humanity's history. The ocean is home to a varied range of habitats and occupies over 70 % of the planet's surface. Biological activity has discovered half of the previously recognized impacts of unique marine natural compounds.<sup>14</sup> The phenomenon of marine ecology is unique. As a result, in order to survive, marine animals must adapt to these harsh conditions. While developing new anticancer treatments, researchers are becoming increasingly interested in marine sources.<sup>15</sup> Prostate cancer is one of the most common cancers affecting men, making its study crucial for early detection and effective treatment. Understanding the disease's progression and risk factors can lead to better screening methods, potentially reducing mortality rates. Research can also contribute to the development of more targeted therapies, improving patient outcomes. Moreover, studying prostate cancer can uncover genetic and environmental influences, aiding in prevention strategies.

## 2. Etiology and risk factors

### 2.1. Diet and obesity

Men in Western nations have a greater incidence of PCa than men in developing/non-Western countries, according to ecological research,<sup>16</sup> showing that nutrition and lifestyle may play a role in PCa epidemiological data. Non-Western males who relocated to Western nations implemented identical lifestyles and PCa risks as those in Western nations, validating this putative link.<sup>17–19</sup> Despite this, no prospective studies linking self-reported dietary patterns of healthful foods and the occurrence of PCa have been published since these ecological descriptions.<sup>20,21</sup> Statins have been shown to reduce the risk of advanced PCa, implying that cholesterol may contribute to the establishment of PCa.<sup>22</sup> Regardless, understanding how diet impacts PCa risk is restricted due to the Western diet's complexity and its interaction/association with good behaviors. Obesity has become a national pandemic in the United States (US), with studies showing that obese persons are more prone to get PCa.<sup>23,24</sup> Obesity and PCa are hypothesized to be linked to greater amounts of free IGF-1, insulin, and estradiol, as well as decreased concentrations of adiponectin and free testosterone.<sup>25</sup> Obesity has been linked to lower blood PSA levels and larger prostates, resulting in fewer prostate biopsies, while there is no clear pathophysiological link between obesity as well PCas.<sup>26</sup>

### 2.2. Inflammation

Chronic inflammation has been linked to the emergence of a lot of

cancers, including PCa, and it might similarly be linked to PCa. Viruses, dietary poisons, hormone imbalances, and physical and chronic stress have all been suggested as possible causes.<sup>25</sup> Intra-prostatic inflammation can produce angiogenesis, cellular turnover, epithelial proliferation, and DNA damage.<sup>27</sup> Those who had at least one inflammation biopsy core had an odds ratio (OR) of 1.78 (95 percent CI 1.04–3.06) for PCa in the PCa Prevention Trial (PCPT) compared to men who had no cores of inflammation in the placebo arm.<sup>28</sup> The association was significantly stronger when considering a diagnosis of high-grade PCa (OR 2.24, 95 percent CI 1.06–4.71).<sup>29</sup>

### 2.3. Medications

As previously stated, HMG-CoA reductase inhibitors have been linked to a lower risk of PCa death after diagnosis.<sup>30</sup> Despite this, the role of statins in prostate carcinogenesis and prevention continues to be a contentious issue. Metformin use has been linked to the same level of optimism as statin use when it comes to PCa outcomes. Metformin has been associated to lower PCa-specific (HR 0.76, 95 percent CI 0.64–0.89 for each extra 6 months of metformin treatment in diabetics) and overall morbidity.<sup>31</sup> According to a meta-analysis and systematic review of observational research evaluating patient clinical outcomes with metformin and PCa, Metformin usage was related with a slightly decreased incidence of biochemical recurrence (HR 0.82, 95 percent CI 0.67–1.01), but not with all-cause death, PCa death, or metastasis.<sup>32</sup>

### 2.4. Genetics

With chromosomal number changes, structural rearrangements, point mutations, and somatic copy number variations, the genetic makeup of PCa is known to be exceedingly complex.<sup>33</sup> PCa is estimated to be caused by mostly inherited genetic factors in 5–10 % of cases.<sup>13</sup> Only a few examples include HPC1, HPC2, HPC20, HPCX, PCAP, and CAPB.<sup>34</sup> Epigenetic markers for PCa, such as miRNA, have been investigated in recent studies. Since the initial study of miRNA and PCa in 2007, more than 30 different miRNAs have been linked to the disease.<sup>35</sup>

## 3. PCa development and hyperplasia

The stroma covers the basal and luminal epithelial cell layers of the growing prostate gland. Basal cells are long, elongated cells that function as a wall among the stroma and the lumen. Luminal epithelial cells are polarized, columnar cells that border the prostate lumen. Different biological markers can be used to identify these cells in terms of disease. Luminal epithelia, for example, exhibits high AR levels and is favorable to cytokeratins eight & Eighteen, as well as NKX3.1 and CD57. AR is expressed in basal epithelial cells at modest levels, along with p63, CD44, cytokeratin 5 and 14, and GSTP1. Neuroendocrine cells produce growth factors, post-mitotic, AR-negative, and neuropeptides for luminal cell development.<sup>36</sup> Eventually, although not the focus of this work, stroma may have a significance in PCa progression and epithelial cell homeostasis,<sup>37,38</sup> multiple critical differentiation regulators, as an instance AR, Forkhead box A1 (FOXA1), and NKX3.1, are required for prostate embryonic and postnatal development. The NKX3.1 transcription factor belongs to the NK homeobox gene family, which controls animal organ development. The earliest prostate-specific gene, NKX3.1, can be found in the prostate as early as embryonic day 15.5.<sup>39</sup> The presence of NKX3.1 in luminal epithelial nuclei corresponds to active morphogenesis at the tip of prostate buds in newborn mice, whereas ductal branching and secretory protein synthesis are reduced when nkx3.1 is genetically deleted.<sup>40</sup> Despite the fact that androgens regulate gene nkx3.1 and that adult mouse castration reduces protein expression, NKX3.1 appears to occur before androgen production, indicating that distinct pathways for expression initiation exist during development.<sup>41</sup> After androgen deprivation early in the development of a mouse embryo, ADT causes lumen involution and luminal epithelial cell death in adult prostate glands,

something that could perhaps be reversed by testosterone replenishment.<sup>42</sup> AR signaling promotes pro-development genes and prostate terminal end buds differentiation, but androgens are not linked to PCa risk, and testosterone use does not increase the risk.<sup>43,44</sup> The variations in androgen function between developing and normal prostates, as well as androgens' role in PCa progression and growth (Fig. 1) are highlighted by these findings. FOXA1 transcription factor influences AR target gene expression throughout the prostate's lifecycle. Deletion of FOXA1 in mice leads to immature epithelial cells and stroma in pubertal ducts. Developmental gene dysregulation promotes abnormal growth and proliferation. PCa should be detected and treated promptly. Identifying PCa origin cell susceptible of tumor induction is one aspect of PCa biology that could provide insight into tumor prognosis and therapy possibilities.<sup>45</sup>

#### 4. Modern treatments for PCa

Most patients can benefit from surgery or radiotherapy when PCa is limited to the adjacent tissues and prostate.<sup>46</sup> For metastatic/advanced disease, androgen restriction therapy is the standard treatment. This approach, however, has become less common in recent years due to the significant risk of resistance.<sup>13</sup> Bilateral orchiectomy is a frequent androgen suppression procedure, but it carries a number of dangers and side effects, including the chance of castration-resistant PCa (CRPC).<sup>47</sup> In affluent countries, this strategy is rarely utilized. Chemical treatments that target gonadotropin-releasing hormone can be used instead of surgical castration. They decrease downstream testosterone synthesis by reducing the production of follicle-stimulating hormone and luteinizing hormone.<sup>48,49</sup> Since 2010, five innovative medications have been approved in the US, with a median life extension of only 3–5 months.<sup>50</sup> Sipuleucel-T, an autologous cellular immunotherapy, is used for less symptomatic disease. Pre- and postdocetaxel therapies include abiraterone acetate (CYP17/androgen biosynthesis inhibitor), enzalutamide (androgen receptor AR inhibitor), and cabazitaxel (microtubule

inhibitor). When other therapies are inadequate, radium-223, a radiopharmaceutical, is used to treat post-docetaxel and docetaxel-ineligible symptomatic bone metastases.<sup>51,52</sup> PSA testing and other technologies for early detection of PCa patients may have resulted in a lower mortality rate. However, people with CRPC still have a high mortality rate.<sup>44</sup> Despite the introduction of novel therapeutic techniques, existing drugs seldom cure this cancer, and survival is usually limited to 4–6 months. Because existing approaches to curative and palliative PCa treatments are linked with high costs and melancholy,<sup>45</sup> PCa chemoprevention ought to be the primary plan for decreasing PCa fatality.<sup>51,53</sup>

#### 5. Chemo-dietary prevention of PCa

One of the first chemoprevention treatments was the long-term use of 5 $\alpha$ -reductase inhibitors like finasteride, a medication approved by the US Food and Drug Administration (FDA). Finasteride inhibits type II and type III 5 $\alpha$ -reductases while having no effect on type I enzymes.<sup>45,46</sup> 5 $\alpha$ -reductase is an enzyme that transforms testosterone to DHT, which stimulates cell development, in individuals with untreated PCa. The PCa Prevention Trial, additionally recognized as the Finasteride Clinical Trial, was a seven-year study that enrolled 18,882 men between the ages of 55 and 65.<sup>49</sup> According to an 18-year follow-up of this study, in the finasteride-treated group, high-grade PCa (Gleason 8–10) was somewhat more prevalent than in the placebo group. Additionally, there was no substantial distinction in as a whole survival or survival following PCa diagnosis between the control and treatment groups.<sup>54</sup> Chau et al.<sup>55</sup> published a research recently looking at serum finasteride concentrations reported in the PCPT trial. Researchers found no link between finasteride levels and PCa risk, but found two single nucleotide polymorphisms in the CYP3A4 enzyme linked to lower plasma levels, while higher levels were associated with other SNPs.<sup>56</sup> In contrast to the PCPT study, the SELECT experiment, which used oral L-selenomethionine (0.2 mg/day) and Vitamin E (-tocopherol, 400 IU/day) in identical demographic

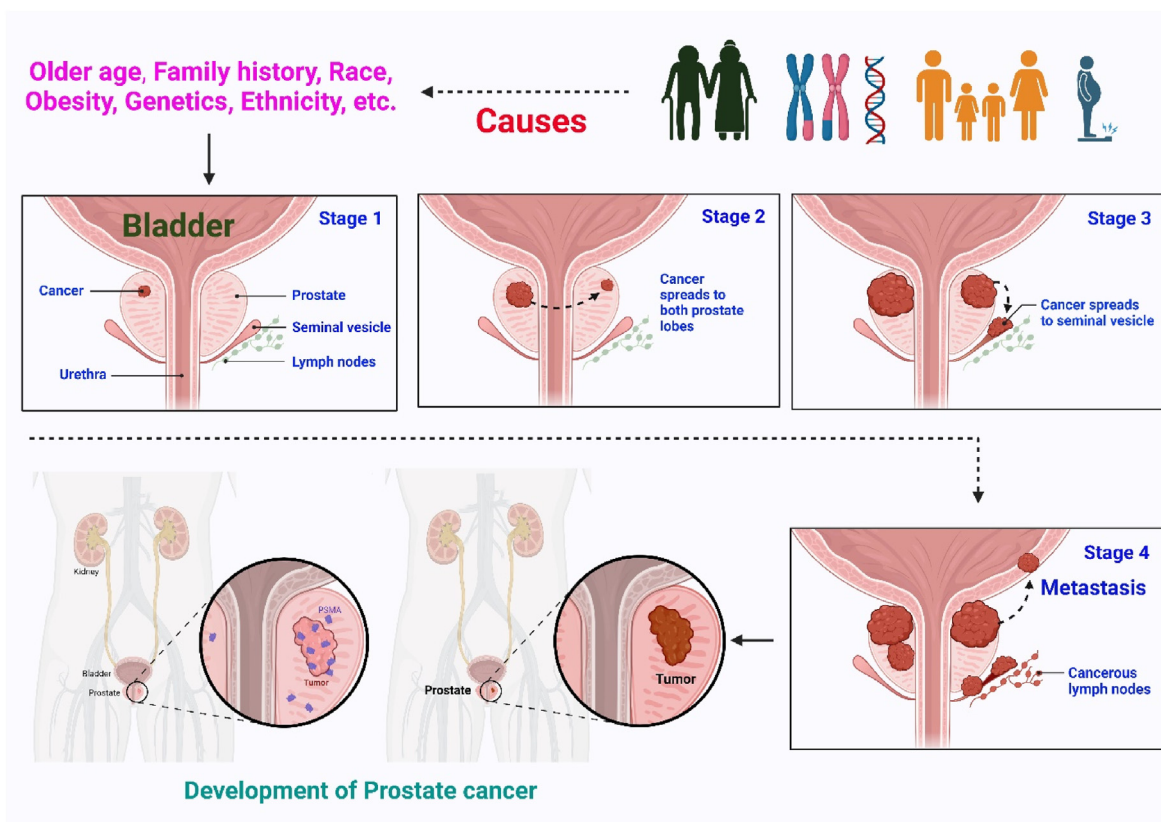


Fig. 1. Causes, stages and progression of PCa.

groups, had a negative outcome.<sup>57</sup> In the SELECT study, selenium supplementation had no effect on PCa risk, whereas vitamin E supplementation had a statistically insignificant effect on PCa risk.<sup>58</sup> The discovery that an exponential growth in 25-hydroxy vitamin D in the blood of the participants reduced PCa incidence was a silver lining in this trial.<sup>59</sup> A new proposal from the PCa treatment and prevention committee suggests that few medicines have been tested in the high-risk or general population groups,<sup>60</sup> patients with PSA4 ng/ml should be given the choice of actively monitoring instead of therapy, emphasizing the relevance of PCa progression prevention techniques. Research on natural-derived substances is needed for chemopreventative therapy for PCa, as 20–30 % of active monitoring patients progress to more aggressive forms.<sup>61</sup>

## 6. Bioactive natural products against PCa

The most promising bioactive natural compounds for the therapy of PCa, as well as their different modes of action, are discussed, as they affect metastasis, invasion, tumor angiogenesis, autophagy, apoptosis, cell cycle control, and cell proliferation (Table 1) (Fig. 2).

### 6.1. Apigenin

Apigenin is a Asteraceae family flavone of plants, including *Anthemis* sp., as well as fruits and vegetables.<sup>153</sup> Apigenin has been shown to inhibit cancer cell growth in a variety of cancer cell lines (lung, liver, colon, and breast).<sup>154</sup> Apigenin (5–40 M) inhibits cell growth and induces death in PC-3 cells, according to another study.<sup>155</sup> Apigenin inhibits cell motility and invasion in PC-3M cells (25 for 16 h) through changing the actin cytoskeleton and decreasing FAK/scr signaling.<sup>156</sup> When given at similar concentrations (20–40 M), apigenin suppresses HDACs class I in both 22Rv1 and PC-3 cells, making it an epigenetic modulator.<sup>157</sup> The generation of ROS causes apoptosis in 22Rv1 cells, which then activates both p53-dependent and p53-independent transcriptional pathways.<sup>158</sup> Apigenin's antiangiogenic properties were as evidenced by C4–2B, LNCaP, and PC-3 cells, with lower VEGF production leading to cancer progression and metastasis reduction.<sup>159</sup> Lastly, Apigenin has been proven in vivo to reduce the development of PC-3 and 22Rv1 tumor xenografts in athymic nude mice,<sup>160</sup> as well as inhibiting cancer progression in TRAMP animals.<sup>161</sup>

### 6.2. Berberine

Isoquinoline alkaloid berberine,<sup>162</sup> found in Berberis plants, is an isoquinoline alkaloid. Berberine reduces DU145, PC-3, and LNCaP cell proliferation and G1 cell cycle arrest in a dosage and time (24–72 h) dependent manner, but has no effect on normal human prostate epithelial cells. Biochemical mechanisms include increased synthesis of Cdk inhibitory proteins (Kip1/p27 and Cip1/p21), increased binding of Cdk inhibitors to Cdks, and inhibition of cyclin-dependent kinase Cdk6, Cdk4, and (Cdk) 2 proteins, as well as cyclins E, D2, and D1. Berberine kills cancer cells by altering the membrane potential of mitochondria, activating poly (ADP-ribose) polymerase, and inducing caspase activation.<sup>163</sup> Ultimately, berberine (at doses of 30 and 50 mg/mL) increases radio sensitivity in human PCa cells via blocking the ROS and MAPK/caspase-3 pathways, as well as reducing the production of HIF-1 $\alpha$  and VEGF.<sup>36</sup>

### 6.3. Capsaicin

Capsaicin is a Capsicum plants derivative vanilloid. Capsaicin inhibits PC-3 cell growth in xenograft models (5 mg/kg) and in vitro (IC<sub>50</sub> of 20 M) by producing ROS, dissipating the inner transmembrane potential of mitochondria, and activating caspase 3.<sup>164</sup> Ceramide buildup, as well as JNK and ERK activation, has been demonstrated cause PC-3 cells apoptosis.<sup>165</sup> At various dosages (100–500), capsaicin increases apoptosis in both androgen-dependent (LNCaP) and resistant (DU-145)

PCa cell lines, and is linked to an boost in Bax, p21, p53 and down-regulation of PSA and AR and decreased proteasome activity.<sup>166</sup>

### 6.4. Curcumin

Curcumin **210**, a diphenylheptanoid derived from *Curcuma longa*, was the first diphenylheptanoid by Inhibiting the EGF-R signaling pathway induces apoptosis in both androgen-resistant and androgen-dependent prostate cancers at dosages ranging from 5 to 50 M.<sup>167</sup> Curcumin promotes apoptosis by interfering with Bcl proteins, producing ROS, and activating mitochondrial stimulation-related pathways. Apoptosis is induced in PC-3 cells by caspase-independent and AIF mechanisms.<sup>168</sup> Curcumin suppresses PCa cell proliferation by downregulating the androgen receptor while activating caspase-dependent apoptosis by upregulating AP-1, NF- $\kappa$ B, CREB, PSA, and cyclin D.<sup>169</sup> Furthermore, PCa cells accumulate in the G1 phase due to proteasome-mediated down-regulation of cyclin E and overexpression of CDKs. Curcumin acts as a chemoprotective medication in early-stage PCa, triggering autophagy and changing Wnt/-catenin pathways.<sup>170</sup> Curcumin likewise suppresses glyoxalases and inhibits histone acetyltransferase, regulating metabolic cellular processes and serving as a histone acetyltransferase blocker.<sup>171</sup> By interfering with cell cytoskeleton architecture and VEGF production, curcumin suppresses PC angiogenesis and metastasis. When given at doses of 5 mg/kg three times a week for four weeks, curcumin decreases invasion and metastasis in DU145 xenografts.<sup>172</sup> In the first in vivo model studied, LNCaP xenograft in nude mice, curcumin inhibited PCa development and reduced metastasis.<sup>173</sup> Liposomal administration, coloaded lipid-based carriers, cellulose nanoparticles, and curcumin-loaded nanospheres have all been shown to improve curcumin transport to cancer cells.<sup>174</sup>

### 6.5. Fisetin

Fisetin is a flavonol with cytotoxic and cytostatic properties in a range of cancer cell lines (pancreatic, ovarian, melanoma, lung, liver, blood, and breast).<sup>175</sup> *Acacia greggii* is one of the plants that contain it. Fisetin promotes G1 cycle arrest in prostate LNCaP cells by downregulating cyclin-dependent kinases and cyclins and initiating both apoptotic pathways caspase-independent and - dependent when administered at 10–60 M for 24 and 48 h.<sup>176</sup> Fisetin inhibits AR production by competing with the AR ligand.<sup>177</sup> Fisetin is an inhibitor of metastasis, migration, and adhesion in highly metastatic PC-3 cells via interfering with the NF- $\kappa$ B pathway and decreasing MMP-9 and MMP-2 expression.<sup>178</sup> In PC-3, DU145, and LNCaP cells, downregulation of NF- $\kappa$ B is associated to an increase in TRAIL-induced apoptosis.<sup>179</sup> Fisetin inhibits the mTOR and PI3K/Akt signaling pathways, causing autophagic cell death.<sup>180</sup> A fisetin treatment (1 mg/animal) twice weekly in the CWR22 Rupsilon1 human xenograft model on Tuesdays reduced tumor progression and PSA levels.<sup>181</sup>

### 6.6. Genistein

Glycine max contains the flavanone genistein **182**. By acting as an inhibitor of tyrosine protein kinase and reducing protein phosphorylation, genistein reduces the proliferation of LNCaP, PC-3, and DU145 PCa cell lines in a dose-dependent manner.<sup>182</sup> Downregulation of mitogen-activated protein kinase 6, cell division cycle 6 (CDC6), DNA topoisomerase II, and survivin, as well as increased regulation of glutathione peroxidase, are all involved in genistein-mediated growth inhibition. In PC-3 cells, IGF-1/IGF-1R signaling pathway suppression has been linked to cell growth suppression.<sup>28,183</sup> As according new research, microRNA regulation is likely to be involved in genistein's apoptotic and antiproliferative activities. As a result, by inhibiting miR-1260b and its targets, sRRP1 and Smad4, genistein promotes apoptosis.<sup>184</sup> Via regulating the expression of miR-34a and HOTAIR, genistein inhibits proliferation of cell in DU145 and PC-3 cells.<sup>185</sup> Apoptosis has also been



**Table 1**  
Anti-PCa bioactive natural compounds.

Sl.No	Compound	Plant	Mechanism of Action	Molecular Pathway	<i>In vitro</i>	<i>In vivo</i>	References
1.	Apigenin	<i>Anthemis</i> sp. ( <i>Asteraceae</i> )	Apigenin induces G1 cell cycle arrest and apoptosis in androgen-dependent and androgen-independent PC cell lines, while inhibiting class I HDACs in PCa-3 and 22Rv1 cells, acting as a modulator of epigenetic events.	Cyclin D1, D2, E, Bax/Bcl-2 ratio, NF- $\kappa$ B, ROS, IGF-IGF-IR, PI3K/Akt HDACs Actin, FAK/scr TIGF-I/IGFBP-3, PI3K/Akt/FoxOGF- $\beta$ , $\downarrow$ VEGF	DU145, LNCaP, PC-3, 22Rv LNCaP, PC-3, C4-2B IGF-1/IGFBP-3, PI3K/Akt/FoxO	22Rv1, PC3 xenografts, TRAMP mice	62,63
2.	Berberine	<i>Berberis</i> sp. ( <i>Berberidaceae</i> )	G1 cycle arrest, Apoptosis. Berberine, given at a dose of 5 mg/kg/day, inhibits tumor growth in LNCaP xenografts in nude mice via lowering AR expression.	Cyclins, CDKs, CDKs inhibitors Bax/Bcl2 ratio, caspases, cytochrome c $\downarrow$ AR expression	DU145, PC-3, LNCaP	LNCaP xenografts	64-67
3.	Capsaicin	<i>Capsicum</i> sp. ( <i>Solanaceae</i> )	Ceramide accumulation and JNK and ERK activation also trigger apoptosis in PC-3 cells.	ROS, JNK, ERK, $\uparrow$ p53, p21, Bax, $\downarrow$ PSA, AR	PC-3, DU145, LNCaP	PC-3 xenografts	68-71
4.	Curcumin	<i>Curcuma longa</i> ( <i>Zingiberaceae</i> )	Curcumin inhibits PCa metastasis and angiogenesis by affecting VEGF synthesis and cell cytoskeleton architecture, thereby promoting cycle arrest and preventing apoptosis in PCa-3 cells.	PC-3, LNCaP	LNCaP xenografts Curcumin-loaded PLGA nanospheres, Cellulose, Nanoparticles, Lipid-based carriers	Cyclins, Cdk, Akt. EGF-R, Bcl, ROS, apoptosis-inducing factor (AIF) $\downarrow$ AR, NF- $\kappa$ B, AP-1, cAMP Response Element-Binding Protein (CREB), PSA.	72-80
5.	Fisetin	<i>Acacia greggii</i> ( <i>Fabaceae</i> )	Cell cycle arrest. Fisetin inhibits the signaling pathways mTOR and PI3K/Akt, resulting in autophagic cell death. In a CWR22 Rupsilon1 human xenograft model, a fisetin injection (1 mg/animal) given 2 times weekly reduced tumor progression and PSA levels.	PC-3, LNCaP	CWR22 Rupsilon1 human xenograft	Cyclins, Cdk, $\downarrow$ AR NF- $\kappa$ B, MMP2, MMP9, mTOR, PI3K/Akt $\downarrow$ PSA	81-85
6.	Genistein	<i>Glycine max</i> ( <i>Fabaceae</i> )	Cycle arrest, growth inhibition. Apoptosis is also linked to a number of pathways, including suppression of proteasomal chymotrypsin-like activity, inactivation of NF- $\kappa$ B, and inhibition of Akt. Decrease Angiogenesis. No metastasis.	DU145, PC-3, NDI, LNCaP, ALVA31, JCA1	PC-3, DU145 xenografts. Lobund-Wistar rats, TRAMP mice Cabazitaxel + genistein Genistein-loaded liposomes	$\uparrow$ p21WAF1, $\downarrow$ Cdk, cyclins, survivin, DNA topoisomerase II, $\downarrow$ IGF-1/IGF-1R NF- $\kappa$ B, Akt, caspases IL-10 MMP-9.	86-95
7.	Ginsenoside Rh2	<i>Panax</i> sp. ( <i>Araliaceae</i> )	$\downarrow$ Cell proliferation, detachment. In a PC-3 human xenograft model in nude mice, oral Rh2 therapy at a dose of 120 mg/kg suppressed cancer cell proliferation, significantly delayed cancer growth, and ultimately increased the rate of apoptosis.	PC-3, LNCaP	PC-3 xenograft	MAP kinases	96-98
8.	Gossypol	<i>Gossypium hirsutum</i> ( <i>Malvaceae</i> )	Due to variations in TGF-beta 1 expression levels, gossypol has been shown to produce G0/G1 cell cycle arrest in PC-3 cells and prostatic cells from people with benign prostatic hyperplasia.	PC-3, BPH DU145 Docetaxel + gossypol	PC-3 xenograft	TGF-1 Bcl-X/Bcl-2, NF- $\kappa$ B, AP1 P53, Bcl-2, Bcl-xL, caspases Beclin1.	28,99-105
9.	Lycopene	<i>Solanum lycopersicum</i> ( <i>Solanaceae</i> )	Cell cycle arrest. $\downarrow$ Cholesterol synthesis. Apoptosis $\downarrow$ Invasion. Apoptosis. Chemoprevention.	LNCaP, PC-3 LNCaP, DU145 LNCaP, PC-3	PC-3 DU145 Xenografts TRAMP mice Clinical trials	Cdk4, cyclins D1, E and Rb PPAR $\gamma$ -LXR $\alpha$ -ABCA1 p21, p27, p53, Bax/Bcl-2, IGF Integrins $\downarrow$ ROS.	106-115
10.	Quercetin	<i>Vitis</i> sp. ( <i>Vitaceae</i> )	Cell cycle arrest. Apoptosis. $\downarrow$ Adhesion, metastasis, $\downarrow$ Angiogenesis, $\downarrow$ Tumor growth	LNCaP, DU145 PC-3	LAPC-4 xenografts	Cyclins, Cdk, Rb, ErbB Bax, Bcl, caspases, IGF, $\downarrow$ fatty acid synthase, $\downarrow$ HSP90, $\downarrow$ AR MMPs.	116-121
11.	Silibinin	<i>Silybum marianum</i> ( <i>Asteraceae</i> )	G1 cycle arrest. G1, G2/M cycle arrest. No invasion, metastasis. $\downarrow$ Tumor growth, apoptosis, angiogenesis. $\downarrow$ Angiogenesis.	LNCaP DU145, 22Rv1 PC-3 PC-3MM2, C4-2B	PC3 xenografts TRAMP	Rb, Cdk, PDEF, $\downarrow$ PSA P21, p27, Wnt/LRp6 HIF1, Wnt/LRp6, cyclins, Cdk, NF- $\kappa$ B, vimentin,	122-134
12.	Sulforaphane	<i>Brassica oleracea</i> ( <i>Brassicaceae</i> )	In both androgen-dependent and androgen-independent cells, SFN has been demonstrated to trigger cell cycle arrest and apoptosis	LNCaP, PC-3 DU145	PC3 xenografts, TRAMP	Methyltransferase, cyclins ROS, caspases, HDAC6, $\downarrow$ AR HIF-1.	135-142

(continued on next page)

Table 1 (continued)

Sl.No	Compound	Plant	Mechanism of Action	Molecular Pathway	<i>In vitro</i>	<i>In vivo</i>	References
13.	Resveratrol	Vitis vinifera	and androgen-refractory PC cell lines. SFN (IC50 of 10 M) causes G2/M phase arrest in DU145 cells. ↓Angiogenesis. Resveratrol is a polyphenol found in grapes that has a number of health benefits, including anti-cancer effects.	PCa models	Target the AR axis	factor 1- $\alpha$ (HIF-1 $\alpha$ ) decreasing $\beta$ -catenin-mediated AR signaling	143–149
14.	Luteolin	López-Lázaro	Luteolin is an anti-inflammatory, anti-cancer, and neuroprotective flavone found in celery, broccoli, parsley, thyme, and rosemary.	Secreted PSA levels	In PCa cells	AR-Hsp90 complex to dissociate the proteasome-ubiquitin	150–152

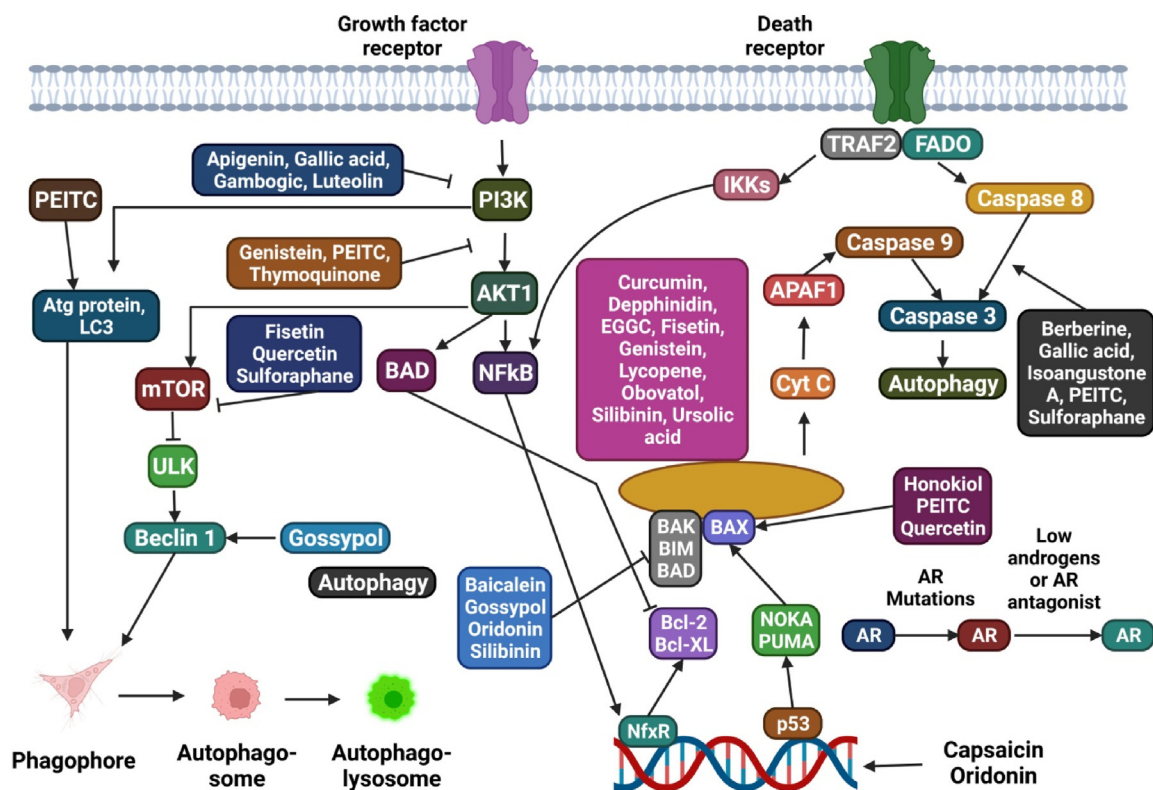


Fig. 2. The mechanism by which bioactive natural compounds interact with PCa cells. Natural substances cause PCa cells to die by activating caspases and mitochondrial-dependent cascades (honokiol), blocking oncogenes (baicalein), and reducing the NF- $\kappa$ B signaling pathway (curcumin).

connected to a number of pathways, including proteasomal chymotrypsin-like activity suppression, NF- $\kappa$ B inactivation, and Akt inhibition.<sup>186</sup> High doses of genistein inhibit growth through regulating the expression of the AR function, which is independent of PSA expression.<sup>39</sup> Genistein, on the other hand, activates mutant forms of AR seen in advanced PC at physiological levels (0.5–5 M).<sup>187</sup> Furthermore, after genistein administration, Many cell migration and adhesion genes (VEGF, uPAR, protease M, MMP-9) are downregulated in PC-3 cells.<sup>30</sup> Cancer stem cells (CSC) are also targeted by genistein and may have an anti-CSC effect, which is significant for reducing metastasis and PC relapse.<sup>188</sup> In DU-145 and PC-3 cells, the epigenetic effects of genistein at 40 M have been shown, where It overturns tumor suppressor gene DNA hypermethylation, resulting in their cancer progression activation and inhibition.<sup>6</sup> Genistein has chemopreventive properties, according to an in vivo investigation. Genistein feeding reduced the incidence of induced prostate-related cancer in L-W rats with spontaneous and generated

metastasizing adenocarcinomas in the prostate-seminal vesicle complex.<sup>50</sup> TRAMP mice were less likely to develop cancer when fed a phytoestrogen-rich diet high in genistein (100, 250, or 500 mg per kilogram).<sup>189</sup> Though oral genistein does not reduce PSA levels in PCa patients, a research shows that 30 mg of synthetic genistein administered every day for 3–6 months reduces PSA levels in the blood.<sup>190</sup> Furthermore, it was observed that combining Cabazitaxel with genistein in the treatment of metastatic-castration-resistant PCa boosted the apoptotic impact.<sup>191</sup> Genistein's clinical efficacy against cancer is limited due to low solubility, poor absorption, and pharmacokinetics. Liposomal vehicle compositions enhance cellular dispersion and proapoptotic efficacy.<sup>192</sup> Finally, researchers did a meta-analysis of the evidence relating soy product consumption to the risk of PCa. According to the findings of this meta-analysis, consumption a lot of non-fermented soy foods can help you from getting PCa.<sup>193,194</sup>

### 6.7. Ginsenosides

Ginsenosides are a type of chemical that only Panax plants produce. Among the bioactive ingredients discovered in root extract of ginseng is ginsenoside Rg3.<sup>195</sup> By boosting the synthesis of cyclin-kinase inhibitors p21 and p27, Rg3 has been demonstrated in the G1 phase to arrest LNCaP cells and eventually initiate a caspase3-mediated death pathway. Rg3 was also demonstrated to modify MAP kinase expression and enhance cell detachment in LNCaP and PC-3 cells, with an IC50 of 8.4 M.<sup>13</sup> Rg3 inhibits PC-3M cell migration and metastasis by interfering with the p38 MAPK pathway, resulting in a decrease in AQP1 expression (a water channel protein implicated in cell migration).<sup>59</sup> Combining Rg3 with several chemotherapeutics (docetaxel, cisplatin, and doxorubicin) reduces PC cell growth more efficiently in PC-3, DU145, and LNCaP cells via suppressing NF- $\kappa$ B activation.<sup>60</sup> Rh2 ginsenoside (-D-glucopyranoside) is a glycoside isolated from the roots of Panax ginseng,<sup>53</sup> that inhibits cell proliferation and causes cell detachment in LNCaP and PC-3 cells with an IC50 of 5.5 M through modifying MAP kinase expression.<sup>195</sup> paclitaxel and Rh2 (0.5–40  $\mu$ M) work together to drastically inhibit LNCaP cell proliferation and tumor formation.<sup>196</sup> In a nude mouse PC-3 human xenograft model, oral Rh2 therapy at a dose of 120 mg/kg suppressed tumor cell proliferation, considerably delayed tumor growth, and finally increased the apoptosis rate.<sup>197</sup> 25-hydroxyprotopanaxadiol (25-OH-PPD) and 20(S)-25-methoxyl-dammarane-3 $\beta$ ,12  $\beta$ ,20-triol (25-OCH<sub>3</sub>-PPD) are also found in Panax ginseng.<sup>198</sup> cyclin D1, E2F1, MDM2, and Cdk 4 and Cdk 2 reduced cell growth-related proteins in PC-3 and LNCaP cells, leading to cell cycle arrest and death. 25-OH-PPD reduced tumor development in PC-3 xenograft tumors dose-dependently.<sup>195</sup>

### 6.8. Gossypol

Gossypol, a polyphenolic aldehyde found in cottonseed,<sup>192</sup> has been demonstrated to have cytotoxic and antiproliferative effects in cell lines of MAT-LyLu and PC cells transplanted in Copenhagen rats. When given at 0.5–4.0 M for 24, 48, or 72 h, gossypol alters TGF1 and Akt signaling in MAT-LyLu cells, changing the regulatory proteins expression like cyclin D1, phospho-Rb, and Cdk4, and eventually inducing G0/G1 cell cycle arrest.<sup>195</sup> Gossypol causes G0/G1 cell cycle arrest in BPH and PC-3 cells, inhibits Bcl-xL/Bcl-2 heterodimerization with proapoptosis molecules, leading to Apoptotic mechanisms that are caspase-dependent and caspase-independent.<sup>199</sup> Gossypol has recently been displayed to induce autophagy in androgen-independent PCa cells with high Bcl-2 levels and resistance to apoptosis, both in vivo and in vitro (PC xenografts), by disrupting the endoplasmic reticulum interaction among Bcl-2/Bcl-xL and Beclin1, releasing the BH3-only pro-autophagic protein Beclin1, which then causes autophag.<sup>196</sup> Gossypol further hinders angiogenesis and metastatic potential (invasion, migration, and adhesion). GP inhibits AP-1 and NF- $\kappa$ B activity in PC-3 cells, resulting in reduced urokinase plasminogen activator and VEGF production as well as chemokine receptor 4 downregulation.<sup>197</sup> Gossypol reduces angiogenesis in human prostate tumor PC-3 xenografts in mice by reducing VEGF receptor 2 kinase activation, which leads to the inhibition of phosphorylation of focal adhesion kinase, AKT kinase, and important intracellular proangiogenic kinases such Src family kinase.<sup>200</sup> In PC-3 cells, a time- and dose-dependent combination of gossypol and docetaxel was revealed to be apoptotic and cytotoxic.<sup>198</sup> Apoptotic and autophagic processes were reported to trigger cell death in DU-145 cells by gossypol (0.5–10 M) and sorafenib (2–20 M), respectively.<sup>201</sup> Finally, it was discovered that giving AT-101 (gossypol) at a dose of 20 mg/day for 21 days to certain men with chemotherapy-naive, castrate-resistant PCa reduced PSA levels.<sup>198</sup>

### 6.9. Lycopene

Lycopene is a carotenoid produced mostly by the tomato *Solanum lycopersicum*.<sup>202</sup> In both LNCaP and DU145 cells, stimulation of the

PPAR-LXR-ABCA1 pathway resulted in a lycopene-mediated reduction in cholesterol synthesis.<sup>194</sup> High dosages of lycopene (16 mg/kg for seven weeks, two times a week) induced apoptosis in PC-3 cells and xenograft models via altering IGF-I, IGF-IR, and IGFBP-3 expression levels.<sup>203</sup> Lycopene inhibits phosphatidylinositol 3-kinase signaling in PC-3 and LNCaP cells, causing cell cycle arrest and apoptosis. It also reduces integrin expression, inhibiting cell motility and invasion.<sup>193,202</sup> Lycopene, on the other hand, is a chemopreventive drug that can help delay or prevent PCa from developing. Lycopene exerts a chemopreventive impact in LNCaP cells by increasing detoxifying proteins, preventing DNA damage, and reducing ROS production and oxidative stress.<sup>189</sup> Lycopene at 4 mg two times daily for one year was demonstrated to reduce or stop the progression of high-grade prostatic intraepithelial neoplasia into PCa, and whole tomato lycopene at 10 mg two times daily for one year lowered PSA velocity in men with PCa.<sup>204,205</sup> Lastly, recent epidemiological studies suggest that lycopene may aid in the prevention of PCa. 5 research investigations indicate a thirty to forty percent risk reduction when eating lots of tomatoes or lycopene, three studies show a 30 % decrease but the findings aren't statically important, and seven research shows no link.<sup>206</sup>

### 6.10. Quercetin

Quercetin is a flavonol that can be found in grapes.<sup>207</sup> In a number of cell lines (blood, bladder, colon, breast, bone, mouth, lung, liver, and esophagus), quercetin has been demonstrated to decrease cell proliferation and activate apoptosis.<sup>208</sup> By interacting with the levels of expression of multiple tumor suppressor genes and oncogenes, quercetin inhibits the growth of PC-3, LNCaP, and DU145 PC cells in a dose-dependent way. Quercetin enhances G2/M cycle arrest in LNCaP by upregulating p21 and inhibiting cyclin B.<sup>209</sup> Lower phosphorylation of MAPK, Akt-1, MAPK kinase 1/2 (MEK1/2), c-Raf, ErbB-3, and ErbB-2, as well as a lowered metastasis rate and drug resistance, appear to be the mechanism of growth suppression in PC-3 cells. Furthermore, quercetin has been linked to a reduction in AR.<sup>210</sup> Through interactions with c-Jun and SP1 proteins at dosages of 5–100 M, quercetin has been found to promote PCa cell lines apoptosis by downregulating heat shock protein and blocking fatty acid synthase.<sup>211</sup> Quercetin in PC-3 cells promotes arrest and death in G2/M cycle by lowering IGF-I, and IGF-II, phosphorylated pRb, cyclin B1, Cdc2/Cdk-1, and increasing caspase-3, Bax, and p21, as well as changing the Bcl-2/Bax ratio.<sup>212</sup> Quercetin promotes TRAIL-induced cytotoxicity by activating caspases, reducing survival, and phosphorylating Akt.<sup>213</sup> When given at a dose of 20 mg/kg/day, in vivo and in vitro investigations in prostate xenograft mouse models reveal quercetin has antiangiogenic effects through interacting with the VEGF-R2-regulated autophagic (AKT/mTOR/P70S6K) pathway.<sup>214</sup>

### 6.11. Silibinin

The flavolignan silibinin, commonly known as silybin, was discovered from the *Silybum marianum* fruits.<sup>215</sup> In LNCaP cells, silibinin has been shown to cause G1 cell cycle arrest as well as intracellular and secretory PSA levels are reduced in a time (12–48 h) and dose- (50–200 M)-dependent manner., with phosphorylation status and modulations of retinoblastoma (Rb) levels, as well as decreased CDK activity.<sup>216</sup> The decrease in PSA in LNCaP cells was found to be caused by downregulation of the epithelium-derived Ets transcription factor (PDEF) and the androgen receptor coactivator.<sup>217</sup> As a lipophilic molecule, silibinin reduces DNA synthesis and mitogenic signaling in DU145 and LNCaP cells, as well as competing in the EGF-erbB1 interaction and lowering DNA synthesis and mitogenic signaling in DU145 and LNCaP cells.<sup>197</sup> In DU145 cells, Silibinin (50–200 M) triggered G1 cell cycle arrest, which was facilitated by a reduction in p27 and p21 expression. Silibinin inhibits Wnt/LRp6 signaling as well as causes apoptosis by suppressing active Stat3, while constitutive NF- $\kappa$ B inactivation makes cells more susceptible to TNF-induced apoptosis.<sup>192</sup> Silibinin causes G2/M and

G1cycle arrest in PC-3 cells by inhibiting the synthesis of CDKs and cyclins as well as the insulin-like growth factor I receptor-mediated signaling pathway in PC-3 cells, causing G2/M and G1cycle arrest<sup>217</sup> at pharmacologically attainable doses (0.02–20 M). PC-3MM2, PC-3, C4–2B LNCaP, as well as DU145 cells have been shown to be resistant to migratory and invasive potential by silibinin<sup>202</sup>. In general, silibinin prevents PC cells from transitioning from epithelial to mesenchymal through interfering with the NF- $\kappa$ B pathway, resulting in downregulation of the transcription factors ZEB1 and SLUG, as well as downregulation of vimentin and MMP2.<sup>46</sup> Silibinin also inhibits osteoclastogenesis in PC cells-induced osteoclastogenesis in high-bone metastatic prostate models.<sup>207</sup> Silibinin has antiproliferative, proapoptotic, and anti-angiogenic effects in xenograft models, increasing Kip1/p27, Cip1/p21, IGFBP-3levels, activating ERK1/2, and decreasing VEGF and Bcl-2 levels in PC-3 tumor xenografts.<sup>218</sup> Although only trace amounts of silibinin were found in prostate tissue, considerable blood silibinin concentrations were found in patients who received a silybin-phytosome (13 g/day) for 14–31 days. The paucity of tissue penetration could be described by silibinin's short half-life, the short length of therapy in this study, or an active process of silibinin removal from the prostate.<sup>208</sup>

### 6.12. Sulforaphane

Sulforaphane (SFN) is an isothiocyanate that occurs naturally in the *Brassica oleracea* plant.<sup>219</sup> In both androgen-refractory and androgen-dependent PC cell lines, SFN has been demonstrated to cause cell cycle apoptosis and arrest. SFN (IC50 of 10 M) causes G2/M phase arrest in DU145 cells,<sup>212</sup> but in LNCaP and PC-3 cells, it causes G1 cell cycle arrest. The antiproliferative effects of SFN are mediated by methyltransferase modulation, which results in a rise in cyclin D2 in LNCaP cells and a decrease in synthesis of protein in PC-3 cells due to reduced phosphorylation of mTOR substrates.<sup>220</sup> Caspases are activated in LNCaP cells, and ROS is produced in PC-3 and DU145 cells, causing intrinsic and extrinsic caspase cascades to be activated.<sup>215</sup> In PC-3, LNCaP, and BPH-1 cells, histone deacetylase 6 is inhibited.<sup>47,221</sup> SFN suppresses prostate cell angiogenesis by inhibiting HIF-1, reducing VEGF production, and inhibiting cell migration in PC-3 and LNCaP cells due to Notch pathway alteration.<sup>214</sup> Oral injection of 7.5 mol per animal per day for twenty-one days reduced tumor volume in PC-3 xenografts in mice that are nude by more than half as a result of a reduction in HDAC activity. Ultimately, broccoli sprouts protected TRAMP rats from PCa.<sup>218</sup>

### 6.13. Resveratrol

Resveratrol is a polyphenol found in grapes that has a number of health benefits, including anti-cancer effects.<sup>212</sup> It was discovered to target the AR axis in PCa models in vitro and in vivo.<sup>47,210,211,219,220</sup> On the one hand, it blocked  $\beta$ -catenin nuclear translocation in LNCaP cells by downregulating hypoxia-inducible factor 1- $\alpha$  (HIF-1 $\alpha$ ), thereby decreasing  $\beta$ -catenin-mediated AR signaling<sup>222</sup>; and it also suppressed interleukin-6 (IL-6)-induced AR transcriptional activity.<sup>223</sup> It assisted the proteasomal degradation of the AR splice variant ARV7 in 22RV1 cells by boosting its polyubiquitination. These findings imply that resveratrol could be used to treat castration-resistant tumors that are ARV7-positive as well as androgen-responsive PCa.<sup>224</sup>

### 6.14. Luteolin

Luteolin is an anti-inflammatory, anti-cancer, and neuroprotective flavone found in broccoli, thyme, parsley, rosemary, and celery.<sup>225,226</sup> It reduced AR mRNA and protein expression in PCa cells, as well as intracellular but also secreted PSA levels, in a dosage and time-dependent manner. It appears to dissociate the AR-Hsp90 complex, allowing the proteasome-ubiquitin route to degrade AR.<sup>227,228</sup>

## 7. Effects on human PCa cells of extracts from specific plants

Recent research on plant families has identified unique bioactive components that could be used as anti-cancer medicines, highlighting their potential as potential chemotherapeutic drugs (Table 2).

### 7.1. Juglandaceae

The majority of members of this family lives in temperate climates. Forests consist of trees and vegetation, with walnuts being nut-producing species. Juglone, a phytochemical, reduces EMT, migration, and invasion in LNCaP-A1 and LNCaP cells.<sup>259</sup> EMT is especially crucial in the course of malignancies because epithelial cancer cells that change to a mesenchymal phenotype are further invasive and thus extra prone to metastasis. Moreover, because bone metastases diminish patient survival rates dramatically,<sup>260</sup> blocking this route could be a promising therapeutic strategy for preventing prostate and other malignancies from spreading.

### 7.2. Moraceae

The Moraceae family includes trees and shrubs that are either deciduous or evergreen and are endemic to tropical and subtropical countries. Many genera are known for their edible fruits, while some are famous for their waxy latex. Members of the Moraceae family have yielded a range of phytochemicals, including chlorogenic acid, flavonoids, ascorbic acid, as well as triterpenoids.<sup>52,282</sup> In PC-3 and LNCaP cells, *Ficus deltoidea* extracts from the deltoidea and angustifolia varieties triggered apoptosis and decreased invasion and migration. Furthermore, through altering the production of vascular endothelial growth factor-A in PC-3 cells, these botanical compounds decreased angiogenesis, a process that promotes tumor formation.<sup>224,283</sup> Similarly, preparations containing significant amounts of ascorbic and chlorogenic acids from *Morus nigra* fruit triggered apoptosis in PC-3 cells and exacerbated cell cycle arrest in the Gap 1 phase.<sup>284</sup> Angiogenesis and uncontrolled proliferation are two characteristics of cancer cells. As a result, stopping either pathway has the potential to stop cancer cells from growing and becoming tumors.<sup>285</sup>

### 7.3. Meliaceae

This family includes shrubs and flowering trees endemic to subtropical and tropical areas. *Azadirachta indica*, also referred as neem, contains nimbolide and terpenoids 28-deoxonimbolide. It inhibits adhesion in LNCaP and PC-3 cells, reduces growth of tumor, and lowers levels of dihydrotestosterone-induced androgen receptors.<sup>286,287</sup> *Trichilia emetic* leaf extracts, on the other hand, have a hormonal effect on PC-3 and LNCaP cells. In PC-3 and LNCaP cells, extracts from the fruit of *Aglaia spectabilis* are cytotoxic.<sup>288</sup> These findings support the idea that Meliaceae plant extracts could be employed in hormone therapy, as well as tumor growth prevention and treatment.<sup>289</sup>

### 7.4. Rutaceae

This is a genus of woody shrubs, flowering and trees native to subtropical and tropical climates worldwide. The most prevalent phytochemicals found in these plants are berberine alkaloids.<sup>290</sup> The bark of *Phellodendron amurense* has the highest concentration of photoberberine. This phytochemical suppressed C4–2B, DU145, LNCaP, and PC-3 cell invasion by reducing NF- $\kappa$ B activity.<sup>291</sup> Vitamin E micelles of berberine inhibited cell proliferation and induced apoptosis in LNCaP and PC-3 cells.<sup>291</sup> Metastasis of PCa has been linked to a poorer survival rate.<sup>287</sup> As a result, it's vital to find compounds that block this pathway.

### 7.5. Bixaceae

Annatto is a red pigment that is used in colors and artworks, is



**Table 2**

The effects of extracts from plants on PCa cells are summarized.

Sl.No	Family	Scientific Name	Mechanism of Action	Part	Type of extract	Dominant phytochemicals	Ref.
1.	Juglandaceae	<i>Juglandaceae</i>	EMT, migration, and invasion are all inhibited by a GSK-3 $\beta$ /snail-dependent mechanism.	Commercial	97 %	Juglone (5-hydroxy-1,4-naphthoquinone)	229,230
2.	Moraceae	<i>Ficus deltoidea</i> L	Stimulation of the intrinsic pathway, modification of the CXCL12-CXCR4 axis to limit both migration and invasion, and manipulation of VEGF-A expression to decrease angiogenesis in PC3 cells	Variety	Crude methanolic extracts: include n-hexane chloroform, and aqueous extracts.	FD1: flavonoid glycosides, furanocoumarin, and chlorophylls FD2: triterpenoids	231–235
3.	Meliaceae	<i>Azadirachta indica</i>	LNCaP lowered prostate-specific antigen levels and dihydrotestosterone-induced androgen receptor. In both cell lines, inhibited the activation of focal adhesion kinase, calreticulin, and integrin $\beta$ 1.	Leaves	Supercritical CO <sub>2</sub> dissolved in DMSO and ethanol	Terpenoids: 28-deoxonimbolide and nimbolide	236
4.	Rutaceae	<i>Phellodendron amurense</i>	Invasion inhibition (through reduced NF- $\kappa$ B activation and its downstream target gene FLIP)	Bark	Commercial (methanolic-palmitate chloride hydrate) Dissolved in water	Protoberberine alkaloid (related to berberine)	59,231–242
5.	Bixaceae	<i>Bixa Orellana</i>	Growth arrest, G1 arrest, and apoptosis are all caused by inhibiting Src and STAT3.	Commercial	Oil	Tocotrienol	243–258
6.	Brassicaceae	<i>Arabidopsis thaliana</i>	Cleaved PARP, a biomarker for OS-induced apoptosis; increased mature CD protein synthesis; p53, a CD transcriptional activator; BAX, a CD downstream effector; and p53, a CD transcriptional activator			Phytoalexin: camalexin	259
7.	Geraniaceae	<i>Biebersteinia multifida</i>	Apoptosis induction and DNA fragmentation	Roots	From 70 % ethanolic extract dissolved into DMSO	Polysaccharides, peptides, alkaloids such as vasicinone, including flavonoids such as apigenin 7-glucosides, luteolin, and tricetin, along with apigenin 7-rutinosides as well as luteolin	260
8.	Lamiaceae	<i>Salvia miltiorrhiza</i> Radix	Apoptosis (increased p21 protein expression and reduced cyclin-dependent kinase 2 (CDK2), CDK4, and cyclin D1 protein levels) as well as cell cycle arrest (increased p21 protein expression and decreased cyclin-dependent kinase 2 (CDK2), CDK4, and cyclin D1 protein levels) were detected.	Commercial	Acetonitrile extract	Lipophilic tanshinones and Hydrophilic phenolic acids, etc.	59,226,241,242,261–271
9.	Apocynaceae	<i>Thevetia peruviana</i> L.	The ability to proliferate and move is hampered. Membrane permeability and DNA fragmentation are two characteristics of apoptosis.	Dried roots, leaves, as well as aerial parts	Methanolic extracts dissolved into DMSO	Polyketide <i>Thevetia</i> flavone as well as the thevetifoline, solanoid, neriifoside cardiac glycosides: peruvosidic acid, peruvoside and neriifolin.	61,225,272–281
10.	Asteraceae	<i>Achillea wilhelmsii</i>	Human telomerase reverse transcriptase inhibition induces apoptosis	leaf and Stem	Hydroalcoholic	sesquiterpene lactones and Flavonoids	224

produced by a large number of members. When PC-3 cells are exposed to a phytochemical tocotrienol, produced from *Bixa orellana* oil, their growth is suppressed. Furthermore, inhibiting Src and STAT3 causes cells to enter in the cell cycle's G1 phase and perish.<sup>283</sup> Deregulation of Src and STAT expression has been associated to a variety of malignancies. Src2 inhibition has also been associated to the suppression of PCa development and metastasis in a mouse model.<sup>292</sup> These findings raise the prospect that these compounds could be employed to treat human cancers.<sup>293</sup>

#### 7.6. Brassicaceae

The family of flowering plants, including broccoli and cabbage, produces isothiocyanate and phytoalexins. Camalexin, found in *Arabidopsis*

*thaliana* extracts, reduces PCa cell viability by increasing oxidative stress and promoting apoptosis proteins.<sup>294</sup> An extract made from the *Brassica juncea* var seed. *Pusa Jaikisan* which includes 3-butenyl isothiocyanate, likewise promotes apoptosis in PC-3 cells. PC-3 cells, revealed a rise in caspase-3 activity in the latter investigation.<sup>295</sup>

#### 7.7. Geraniaceae

These are dicotyledon-flowering shrubs that are indigenous to temperate areastricetin, apigenin 7-rutinoside, and Apigenin 7-glucosides, luteolin, and luteolin are polysaccharides, peptides, alkaloids, and flavonoids discovered in ethanolic extracts of *Biebersteinia multifida* roots. DNA fragmentation and death are enhanced when DU145 and PC-3 cells are exposed to this extract.<sup>296</sup> While fragmentation of DNA can

happen by accident, it is a defining feature of apoptosis and thus a prospective chemotherapeutic target.<sup>297</sup>

### 7.8. Lamiaceae

Flowering aromatics that are endemic to temperate areas all over the world belong to this family of plants. The majority of the plants are herbs, both perennial and annual, with fragrant leaves and blooms. Triterpenoids, lipophilic tanshinones, and Phenolic acids are all members of this family. *Salvia miltiorrhiza*, and *Salvia miltiorrhiza*. All displayed cytotoxic effects on LNCaP, PC-3, or DU145 PCa cells, producing apoptosis and cell cycle arrest through a number of methods.<sup>28,35,297,298</sup> PC-3 cells treated with *Scutellaria altissima* extract, primarily scutellarin, exhibit enhanced GAP2/mitotic phase entry due to increased production of cyclin B1 and Cdc2.<sup>299</sup> *Salvia miltiorrhiza* acetonitrile extract causes cell cycle arrest in PC-3 cells by increasing p21 expression and reducing cyclin-dependent kinases 2 through 4 and cyclin D1.<sup>300</sup> The herb has been found to have hormonal effects on PC-3 and LNCaP cancer cells, suggesting potential use in PCa hormone therapy.<sup>301,302</sup>

### 7.9. Apocynaceae

*Thevetia peruviana*, a genus of tropical and subtropical plants, contains flavones that inhibit cell proliferation, motility, and DNA fragmentation in HTB-81 PCa adenocarcinoma cells, while increasing membrane permeability.<sup>303</sup> *Biebersteinia multifida* is a Geraniaceae family member, that also includes flavonoids, causes DNA fragmentation.<sup>304</sup> Flavonoids may be the reason for DNA fragmentation and subsequent death in PCa cells.<sup>305</sup>

### 7.10. Asteraceae

Flowering herbs, shrubs, and trees from all over the world belong to this family. Many of the species have previously been used as decorative or food plants. Phytochemicals derived from this plant family include flavonoids, sesquiterpenes, and phenols. Anti-proliferative and cytotoxic activities of extracts from *Achillea wilhelmsii*, and *Melampodium leucanthum*, *Verbesina virginica*, against PC-3 as well as DU145 PCa cell lines.<sup>306-308</sup> *Melampodium leucanthum*, containing tricyclic sesquiterpenes and germacranolide sesquiterpene lactones, causes PC-3 and DU145 cells to stop dividing during the GAP2/mitotic phase, leading to an abnormal mitotic spindle.<sup>309</sup> This family appears to have a number of possible chemotherapeutic pathways, making it a promising candidate for further study.

## 8. Marine bioactive products having potential for PCa treatment

### 8.1. Marine bacteria

Microorganisms that live in the marine have a special source of anticancer medicines. Scientists have recently become interested in marine microorganisms in order to improve the development of these medications. *Scytonemin*, *manoalide*, topsentins, and scytonemin are anti-inflammatory compounds found in sea microorganisms.<sup>307,308</sup> Certain substances such as Kahalalide F, have been shown to be cytotoxic to cancer cell lines like PCa-3, but have no effect on hormone-sensitive LNCaP cells in vitro.<sup>300,310</sup> KF has anticancer properties in solid PCa, with clinical trials showing partial responses in one patient and good health in five. It can be safely given as a 1-h infusion on five days in a row once every three weeks.<sup>311,312</sup>

### 8.2. Marine fungi

Marine fungi contain biologically active metabolites, but few researches have explored their effects on androgen-independent cell growth. The South China Sea metabolite 1386A showed inhibitory

concentrations of 25.31, 8.62, and 4.79 mol/L.<sup>313</sup> In diseases like PCa, this activity could be used as a medicine or a food supplement.<sup>314</sup> Marine gut fungus known as aspochalasins that is found in *Ligia oceanica*'s gut, have been identified as bioactive compounds with cytotoxic, anti-HIV, anti-TNF-alpha, and melanogenesis inhibitory effects.<sup>315-317</sup> The PCa PC-3 cell line was tested for cytotoxicity using the MTT method. At IC50 values of 30.4  $\mu$ M, apochalasin V was shown to have just minor activity.<sup>318</sup>

### 8.3. Marine sponges

In marine sponges, alkaloids can be discovered in large quantities. Rhizochalin is one such bioactive molecule obtained from *Rhizochalina incrustata* that is a marine sponge. Rhizochalin reduced promoted apoptosis and autophagy in human castration-resistant PCa cells.<sup>319</sup> Rhizochalinin, a cytotoxic compound derived from rhizochalin, induces apoptosis and cell cycle arrest in human PCa cell lines at low concentrations, with aglycones being more cytotoxic.<sup>320</sup> Functional investigations revealed Rhiz's anti-migratory effect on PC-3 cells, validated by Western blot analysis, and pro-survival effects in Rhiz-treated PCa cells, suggesting a possible resistance mechanism.<sup>321,322</sup> In addition, halichonadiamines (HCA), which are made from the marine sponge *Haliclona* spp. ethanol extracts, have a substantial cytotoxic effect on PC-3 cells, at 100 M with 50 % viability.<sup>323</sup> HCA treatment inhibited PRL-3 cell upregulation, enhanced E-cadherin expression, and suppressed N-cadherin expression, while Latrunculin A, a macrolide from *Negombata magnifica*, showed anti-invasive properties against PC-3 cells.<sup>324</sup> Halichondramide is a trisoxazole-containing macrolide that affects the transcriptional and translational levels of PCa indicators such as MMP2, E-cadherin, MMP9, and N-cadherin.<sup>321</sup> PCa biomarkers that show the epithelial to EMT represent the metastatic potential of the disease.<sup>323</sup> A macrocyclic lactone from *Spongia* sp. has been found to trigger cell death and apoptosis in DU-145 cells without the need for caspases.<sup>325</sup> Spongistatin 1 targets microtubular complex and MCL-1 to upregulate BIM, a pro-apoptotic BCL-2 family member in PCa, regulating Apoptotic signaling pathways.<sup>323</sup> Under hypoxic conditions, furospinosulin-1, a furanosesterterpene derived from marine sponges, inhibits the proliferation of DU-145 cells.<sup>326</sup> By deactivating HIF-1, sodwanone and yardenones generated by *Axinella* sp. inhibited PC-3 cells.<sup>327</sup> Niphatenone B, a natural chemical from *Niphates digitalis*, induces castration-recurrent PCa by promoting LNCaP cell proliferation, preventing AR assistance for specific targets, and binding to AR N-terminus domain activators.<sup>328</sup> Ultimately, Agelasin B was separated using *Agelas clathrodes* is a marine sponge. PC-3 cells have been demonstrated to have a high viability rate. It promotes DNA breakage and significantly reduces the Ca<sup>2+</sup> concentration in these cells.<sup>326</sup>

### 8.4. Marine Diatoms

Diatoms have few natural bioactive chemicals, but Fucoxanthin, a marine substance found in *Sargassum* sp., is used to treat PCa by inhibiting LNCaP cells.<sup>329</sup> Fucoxanthin, a naturally occurring compound, has been shown to be generally safe in treating PCa by inhibiting the growth of GADD45A and G1 cells.<sup>330</sup> White Leghorn fed brown seaweed *F. serratus* deacetylated fucoxanthin in intestinal lumen, transmitting it via circulation, resulting in fucoxanthinol, a major carotenoids found in egg yolks.<sup>331</sup> In a study on the biotransformation of fucoxanthinol in ICR mice, Asai et al found an unknown metabolite in the marine tunicate *Amaroucium pliciferum* that had previously been identified as amarouciaxanthin A. Fucoxanthin, fucoxanthinol, and amarouciaxanthin A had 50 % inhibitory doses of 3.0, 2.0, and 4.6  $\mu$ M,<sup>332</sup> respectively, and Fucoxanthinol with amarouciaxanthin are both antioxidants. PC-3 cell viability was lowered. However, there is a scarcity of research on the subject.

### 8.5. Holothurians

Holothurians (sea cucumbers) are marine crustaceans that have been utilized for thousands of years in traditional Asian medicine.<sup>333</sup> Fronsoside A (FrA) is a triterpene glycoside found in an extract of the edible sea cucumber *Cucumaria frondosa*. In human PCa cells, including cell types resistant to current therapy, the FrA molecule exhibited high efficacy and low toxicity. It has a one-of-a-kind set of features, including apoptosis induction, cell cycle arrest, pro-survival autophagy regulation, and immunological modulatory effects.<sup>334</sup> 12-MTA suppressed PCa cell proliferation and induced apoptosis in PC-3 cells, potentially providing a unique complementary therapy for PCa by preventing 5-HETE synthesis<sup>251</sup>

### 8.6. Cyanobacteria

Cyanobacterium, a diverse bacterial phylum, was found in the marine cyanobacteria *Nostoc* spp., which produces an anticancer chemical called Cryptophycin 52.<sup>252</sup> A novel cyclic depsipeptide was discovered in *L. majuscula*, a marine cyanobacterium from Singapore's Pulau Hantu Besar. PC3 cells were tested with Lagunamide C, which has an IC50 of 2.6 nM. It has anti-malarial effects as well.<sup>334</sup> *Dolabella auricularia* is a bacterial species that produces cytotoxic peptides, including Dolastatin 10 and its synthetic analogs, which cause cell cycle arrest during the G2/M phase.<sup>335</sup> Marine cyanobacteria compounds, such as C-phyco-cyanin, have anticancer effects. Combining 10 % of topotecan with C-PC was found to be more effective in killing cancer cells, with enhanced caspase-9 and caspase-3 activity observed.<sup>336</sup> The BCL-2 protein family is a crucial regulator in PCa that can cause apoptosis or cell death. Cryptophycin 52 increases phosphorylation of BCL-2 and BCL-xL in a number of PCa cell lines, including DU-145, PC-3, and LNCaP.<sup>337</sup> Iejimalide B is active in LNCaP and PC-3 cell lines at nanomolar doses, but its effects

vary significantly, with less than 30 nM causing cell cycle arrest.<sup>338</sup>

## 9. Numerous molecules play a role in the beginning or progression of PCa to CRPC

Many molecules have been implicated in the initiation or progression of PCa to CRPC, and many of these can be inhibited in vivo (Fig. 3). Understanding that not all cell-specific Myc targets are created equal could lead to the discovery of sites that effectively limit Myc activity in PCa initiation.<sup>339</sup> Overall, our understanding of PCa progression to CRPC much outnumbers our understanding of disease etiology. It is certain that PCa treatments will gradually transition to ward precision medicine as whole genome sequencing becomes more common. The ability to pinpoint which mutations and genetic abnormalities are present in a patient's tumor could pave the path for more efficient CRPC treatment. Technological advancements require understanding vast data to develop effective treatments for PCa. Whole genome research can widen the molecular spectrum, but complete understanding of biological circumstances and mutational cross-talk is impossible.<sup>340</sup>

## 10. Conclusion and future perspectives

Using phytochemicals to treat PCa has a number of advantages. Natural products, for starters, are frequently safe, well-liked, and cost-effective. In vivo and in vitro, they have anti-tumor features such as growth anti-invasive, suppression, pro-death, as well as anti-angiogenic effects. However, nutraceutical intake has been linked to chemoprevention and PSA reduction rather than tumor eradication, suggesting that these potential pleiotropic effects in PCa patients have only been partially verified. As a result, more clinical trials targeted at confirming nutraceutical efficacy in humans are directly required.<sup>341</sup> Academics in the US and worldwide have recognized the resistance of marine chemicals

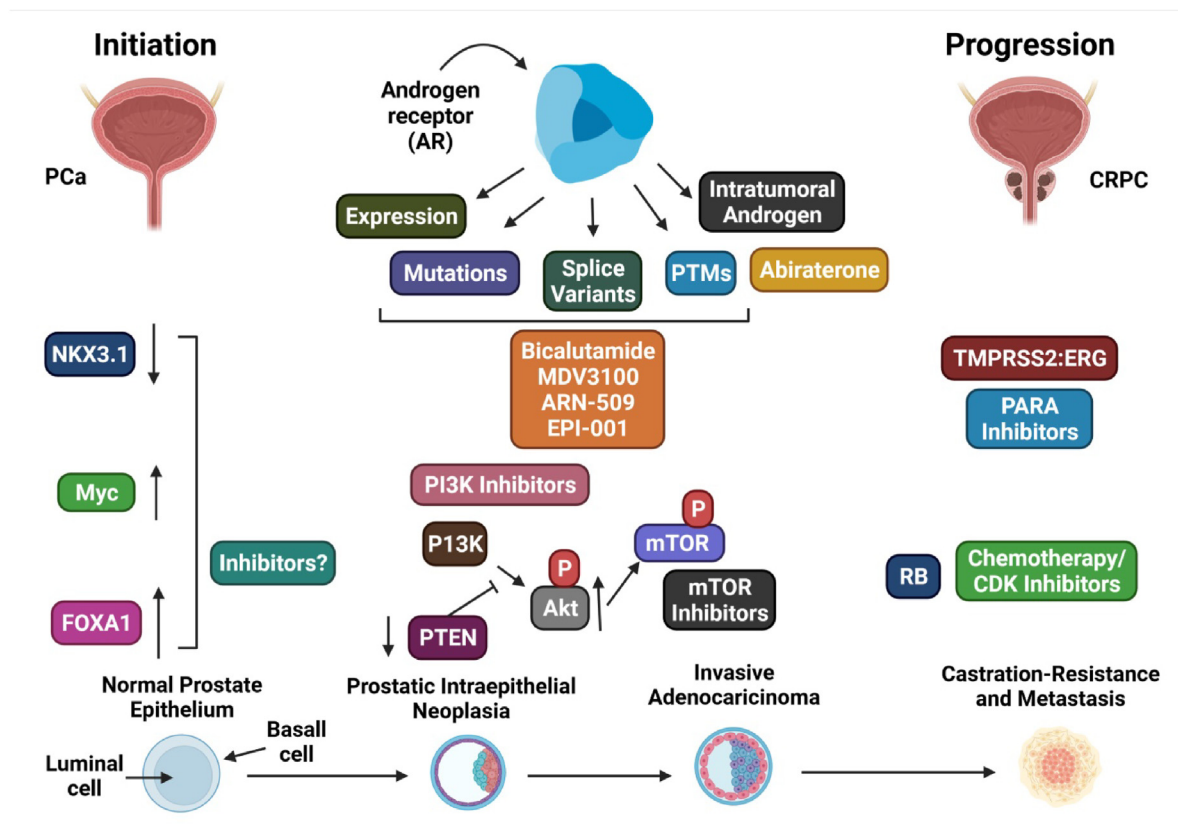


Fig. 3. Molecules linked to the onset and progression of PCa. Several signaling pathways are deregulated with the onset of PCa and progression to castration resistant PCa.

against PCa. The direct effects of marine chemicals on tumor cells through enhancing host immune function are being investigated in the investigation of anti-PCa effects of marine compounds. To summarize, marine chemicals offer a lot of potential as anticancer drugs. PCa has become more prevalent in recent years. Medicinal compounds from land-grown animals and plants face challenges in collection due to duplicates and resource competition, especially for endangered species. Marine sources could help with PCa research. However, very little research has been done on this subject. The species used to assess the anticancer properties of marine compounds are a small part of the ocean's millions of species. There is a lot of maritime flora in nature, and it has been used to extract various anticancer bioactive compounds.<sup>231</sup> This study looked at a number of sea-derived bioactive natural compounds and molecules that have been linked to PCa. Both natural and marine chemicals have the potential to improve PCa patients' quality of life. To develop novel anti-PCa therapies, more study on these substances is required.

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**Konatham Teja Kumar Reddy:** Writing – review & editing. **Karthickeyan Krishnan:** Writing – review & editing, Formal analysis, Conceptualization. **Palani Shanmugasundaram:** Methodology, Investigation, Data curation, Conceptualization. **C. Ronald Darwin:** Validation. **Balaji Pandian:** Writing – review & editing. **Saravanan Govindaraj:** Formal analysis. **Priyanga Jaganath:** Validation. **Sridevi Ganesan:** Writing – review & editing.

### Declaration of competing interest

The authors declare that there are no conflicts of interest.

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### References

- Global Burden of Disease Cancer C, et al. Global, regional, and national cancer incidence, mortality, years of life lost, years lived with disability, and disability-adjusted life-years for 32 cancer groups, 1990 to 2015: a systematic analysis for the global burden of disease study. *JAMA Oncol.* 2017;3(4):524–548.
- Stephenson AJ, et al. Preoperative nomogram predicting the 10-year probability of prostate cancer recurrence after radical prostatectomy. *J Natl Cancer Inst.* 2006; 98(10):715–717.
- Perlmutter MA, Lepor H. Androgen deprivation therapy in the treatment of advanced prostate cancer. *Rev Urol.* 2007;9(Suppl 1):S3–S8. Suppl 1.
- Thomas TS, Pachynski RK. Treatment of advanced prostate cancer. *Mo Med.* 2018; 115(2):156–161.
- Sweeney CJ, et al. Chemohormonal therapy in metastatic hormone-sensitive prostate cancer. *N Engl J Med.* 2015;373(8):737–746.
- James ND, et al. Addition of docetaxel, zoledronic acid, or both to first-line long-term hormone therapy in prostate cancer (STAMPEDE): survival results from an adaptive, multiarm, multistage, platform randomised controlled trial. *Lancet.* 2016; 387(10024):1163–1177.
- Justilien V, Fields AP. Molecular pathways: novel approaches for improved therapeutic targeting of Hedgehog signaling in cancer stem cells. *Clin Cancer Res.* 2015;21(3):505–513.
- Teo MY, Rathkopf DE, Kantoff P. Treatment of advanced prostate cancer. *Annu Rev Med.* 2019;70:479–499.
- Cathomas R, et al. Management of bone metastases in patients with castration-resistant prostate cancer. *Urol Int.* 2014;92(4):377–386.
- Salehi B, et al. Phytochemicals in prostate cancer: from bioactive molecules to upcoming therapeutic agents. *Nutrients.* 2019;11(7).
- Kallifatidis G, Hoy JJ, Lokeshwar BL. Bioactive natural products for chemoprevention and treatment of castration-resistant prostate cancer. *Semin Cancer Biol.* 2016;40–41:160–169.
- Moselhy J, et al. Natural products that target cancer stem cells. *Anticancer Res.* 2015;35(11):5773–5788.
- Fontana F, et al. Natural compounds in prostate cancer prevention and treatment: mechanisms of action and molecular targets. *Cells.* 2020;9(2).
- Kiuru P, et al. Exploring marine resources for bioactive compounds. *Planta Med.* 2014;80(14):1234–1246.
- Müller M, et al. Biochemistry and evolution of anaerobic energy metabolism in eukaryotes. *Microbiol Mol Biol Rev.* 2012;76(2):444–495.
- Center MM, et al. International variation in prostate cancer incidence and mortality rates. *Eur Urol.* 2012;61(6):1079–1092.
- Nipun TS, et al. Active site-specific quantum tunneling of hACE2 receptor to assess its complexing poses with selective bioactive compounds in co-suppressing SARS-CoV-2 influx and subsequent cardiac injury. *J Adv Vet Anim Res.* 2021;8(4): 540–556.
- Kumari R, et al. Repurposing of the herbals as immune-boosters in the prevention and management of COVID-19: a review. *J Pure Appl Microbiol.* 2021;15(1).
- Pervin Z, Hassan MM. Synergistic therapeutic actions of antimicrobial peptides to treat multidrug-resistant bacterial infection. *Rev Med Microbiol.* 2020;32:83–89.
- Ahdoot M, et al. Contemporary treatments in prostate cancer focal therapy. *Curr Opin Oncol.* 2019;31(3):200–206.
- Artibani W, et al. Management of biochemical recurrence after primary curative treatment for prostate cancer: a review. *Urol Int.* 2018;100(3):251–262.
- Bayne CE, et al. Treatment of the primary tumor in metastatic prostate cancer: current concepts and future perspectives. *Eur Urol.* 2016;69(5):775–787.
- Belderbos BPS, et al. Novel treatment options in the management of metastatic castration-naïve prostate cancer; which treatment modality to choose? *Ann Oncol.* 2019;30(10):1591–1600.
- Bilusic M, Madan RA, Gulley JL. Immunotherapy of prostate cancer: facts and hopes. *Clin Cancer Res.* 2017;23(22):6764–6770.
- Brawley S, Mohan R, Nein CD. Localized prostate cancer: treatment options. *Am Fam Physician.* 2018;97(12):798–805.
- C DEN, et al. Castration-resistance prostate cancer: what is in the pipeline? *Minerva Urol Nefrol.* 2018;70(1):22–41.
- Cackowski FC, et al. Evolution of disparities in prostate cancer treatment: is this a new normal? *Am Soc Clin Oncol Educ Book.* 2021;41:1–12.
- Chen FK, de Castro Abreu AL, Palmer SL. Utility of ultrasound in the diagnosis, treatment, and follow-up of prostate cancer: state of the art. *J Nucl Med.* 2016; 57(Suppl 3):13s–18s.
- Clarke JM, Armstrong AJ. Novel therapies for the treatment of advanced prostate cancer. *Curr Treat Options Oncol.* 2013;14(1):109–126.
- Denmeade SR, Isaacs JT. A history of prostate cancer treatment. *Nat Rev Cancer.* 2002;2(5):389–396.
- Derleth CL, Yu EY. Targeted therapy in the treatment of castration-resistant prostate cancer. *Oncology (Williston Park).* 2013;27(7):620–628.
- Emmett L, et al. Lutetium (177) PSMA radionuclide therapy for men with prostate cancer: a review of the current literature and discussion of practical aspects of therapy. *J Med Radiat Sci.* 2017;64(1):52–60.
- Gamat M, McNeel DG. Androgen deprivation and immunotherapy for the treatment of prostate cancer. *Endocr Relat Cancer.* 2017;24(12):T297–t310.
- Haberkorn U, et al. New strategies in prostate cancer: prostate-specific membrane antigen (PSMA) ligands for diagnosis and therapy. *Clin Cancer Res.* 2016;22(1): 9–15.
- Foster CC, Weichselbaum RR, Pitroda SP. Oligometastatic prostate cancer: reality or figment of imagination? *Cancer.* 2019;125(3):340–352.
- Heidegger I, et al. Novel therapeutic approaches for the treatment of castration-resistant prostate cancer. *J Steroid Biochem Mol Biol.* 2013;138:248–256.
- Janiczek M, et al. Immunotherapy as a promising treatment for prostate cancer: a systematic review. *J Immunol Res.* 2017;2017:4861570.
- Kaiser A, et al. The evolving role of diet in prostate cancer risk and progression. *Curr Opin Oncol.* 2019;31(3):222–229.
- Katz A. What happened? Sexual consequences of prostate cancer and its treatment. *Can Fam Physician.* 2005;51(7):977–982.
- Komura K, et al. Current treatment strategies for advanced prostate cancer. *Int J Urol.* 2018;25(3):220–231.
- Koo KC, Dasgupta P. Treatment of oligometastatic hormone-sensitive prostate cancer: a comprehensive review. *Yonsei Med J.* 2018;59(5):567–579.
- Kuyu H, et al. Recent advances in the treatment of prostate cancer. *Ann Oncol.* 1999; 10(8):891–898.
- Lilleby W, et al. [Treatment of hormone-resistant prostate cancer]. *Tidsskr Nor Laegeforen.* 2006;126(21):2798–2801.
- Lodeizen O, et al. Ablation energies for focal treatment of prostate cancer. *World J Urol.* 2019;37(3):409–418.
- Logothetis CJ, et al. Molecular classification of prostate cancer progression: foundation for marker-driven treatment of prostate cancer. *Cancer Discov.* 2013; 3(8):849–861.
- Maffioli L, et al. New radiopharmaceutical agents for the treatment of castration-resistant prostate cancer. *Q J Nucl Med Mol Imaging.* 2015;59(4):420–438.



47. Maráz A, et al. [Therapeutic sequences in the treatment of advanced/metastatic prostate cancer]. *Magy Onkol.* 2020;64(3):263–272.
48. McKay RR, et al. Recent advances in the management of high-risk localized prostate cancer: local therapy, systemic therapy, and biomarkers to guide treatment decisions. *Am Soc Clin Oncol Educ Book.* 2020;40:1–12.
49. Miyahira AK, et al. Prostate cancer research: the next generation; report from the 2019 coffey-holden prostate cancer academy meeting. *Prostate.* 2020;80(2):113–132.
50. Mulders PF, et al. Targeted treatment of metastatic castration-resistant prostate cancer with sipuleucel-T immunotherapy. *Cancer Immunol Immunother.* 2015;64(6):655–663.
51. Nader R, El Amm J, Aragon-Ching JB. Role of chemotherapy in prostate cancer. *Asian J Androl.* 2018;20(3):221–229.
52. Nussbaum N, et al. Patient experience in the treatment of metastatic castration-resistant prostate cancer: state of the science. *Prostate Cancer Prostatic Dis.* 2016;19(2):111–121.
53. Parker C, et al. Prostate cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* 2020;31(9):1119–1134.
54. Rosellini M, et al. Treating prostate cancer by antibody-drug conjugates. *Int J Mol Sci.* 2021;22(4).
55. Serrano NA, Anscher MS. Favorable vs unfavorable intermediate-risk prostate cancer: a review of the new classification system and its impact on treatment recommendations. *Oncology (Williston Park).* 2016;30(3):229–236.
56. Shore ND, et al. Building on sipuleucel-T for immunologic treatment of castration-resistant prostate cancer. *Cancer Control.* 2013;20(1):7–16.
57. Spratt DE, et al. Management of biochemically recurrent prostate cancer: ensuring the right treatment of the right patient at the right time. *Am Soc Clin Oncol Educ Book.* 2018;38:355–362.
58. Tsujino T, et al. CRISPR screen contributes to novel target discovery in prostate cancer. *Int J Mol Sci.* 2021;22(23).
59. Wadia R, Petrylak DP. New developments in the treatment of castration resistant prostate cancer. *Asian J Androl.* 2014;16(4):555–560.
60. Wallis CJD, et al. Surgery versus radiotherapy for clinically-localized prostate cancer: a systematic review and meta-analysis. *Eur Urol.* 2016;70(1):21–30.
61. Williams SB, et al. Risk of hospitalisation after primary treatment for prostate cancer. *BJU Int.* 2017;120(1):48–55.
62. Abeygunasekera AM, et al. Clinicopathological characteristics and primary treatment of prostate cancer in a urology unit of Sri Lanka. *J Cancer Res Ther.* 2015;11(4):780–785.
63. Acar O, Esen T, Lack NA. New therapeutics to treat castrate-resistant prostate cancer. *Sci World J.* 2013;2013:379641.
64. Akaza H. Current status and prospects of androgen depletion therapy for prostate cancer. *Best Pract Res Clin Endocrinol Metabol.* 2008;22(2):293–302.
65. Al Otaibi M, et al. Role of repeated biopsy of the prostate in predicting disease progression in patients with prostate cancer on active surveillance. *Cancer.* 2008;113(2):286–292.
66. Allen GW, et al. Management of prostate cancer recurrences after radiation therapy-brachytherapy as a salvage option. *Cancer.* 2007;110(7):1405–1416.
67. Altwaijry N, Somani S, Dufès C. Targeted nonviral gene therapy in prostate cancer. *Int J Nanomed.* 2018;13:5753–5767.
68. Aneja S, Pratiwadi RR, Yu JB. Hypofractionated radiation therapy for prostate cancer: risks and potential benefits in a fiscally conservative health care system. *Oncology (Williston Park).* 2012;26(6):512–518.
69. Aparicio A, Tzelepi V. Neuroendocrine (small-cell) carcinomas: why they teach us essential lessons about prostate cancer. *Oncology (Williston Park).* 2014;28(10):831–838.
70. Aragon-Ching JB, Williams KM, Gulley JL. Impact of androgen-deprivation therapy on the immune system: implications for combination therapy of prostate cancer. *Front Biosci.* 2007;12:4957–4971.
71. Arlen PM, et al. Promising novel immunotherapies and combinations for prostate cancer. *Future Oncol.* 2009;5(2):187–196.
72. Auran-Gomez AM, Scarpa RM, Chin J. High-intensity focused ultrasound and cryotherapy as salvage treatment in local radio-recurrent prostate cancer. *Urol Int.* 2012;89(4):373–379.
73. Azuma H, Katsuoka Y. [Treatment for locally advanced prostate cancer: value of surgery]. *Hinyokika Kyo.* 2006;52(6):459–467.
74. Ba Dziul D, et al. Mathematical prostate cancer evolution: effect of immunotherapy based on controlled vaccination strategy. *Comput Math Methods Med.* 2020;2020:7970265.
75. Baker BR, et al. Use of stereotactic body radiotherapy for prostate cancer in the United States from 2004 through 2012. *Cancer.* 2016;122(14):2234–2241.
76. Baldwin LM, et al. Treatment of early-stage prostate cancer among rural and urban patients. *Cancer.* 2013;119(16):3067–3075.
77. Bannuru RR, et al. Comparative evaluation of radiation treatments for clinically localized prostate cancer: an updated systematic review. *Ann Intern Med.* 2011;155(3):171–178.
78. Bardia A, et al. Anti-inflammatory drugs, antioxidants, and prostate cancer prevention. *Curr Opin Pharmacol.* 2009;9(4):419–426.
79. Baronzio G, Gramaglia A, Fiorentini G. Review. Current role and future perspectives of hyperthermia for prostate cancer treatment. *In Vivo.* 2009;23(1):143–146.
80. Barry Delongchamps N. Prostate cancer: review in 2014. *Diagn Interv Imag.* 2014;95(7–8):739–742.
81. Bass EJ, Ahmed HU. Focal therapy in prostate cancer: a review of seven common controversies. *Cancer Treat Rev.* 2016;51:27–34.
82. Bates M, et al. YB-1: the key to personalised prostate cancer management? *Cancer Lett.* 2020;490:66–75.
83. Baumgart SJ, Haendler B. Exploiting epigenetic alterations in prostate cancer. *Int J Mol Sci.* 2017;18(5).
84. Beesley LJ, et al. Individual and population comparisons of surgery and radiotherapy outcomes in prostate cancer using bayesian multistate models. *JAMA Netw Open.* 2019;2(2):e187765.
85. Bergman J, et al. Partnership and outcomes in men with prostate cancer. *Cancer.* 2009;115(20):4688–4694.
86. Bernard B, et al. Approach to oligometastatic prostate cancer. *Am Soc Clin Oncol Educ Book.* 2016;35:119–129.
87. Berthelet E, et al. Consistency in electronic portal imaging registration in prostate cancer radiation treatment verification. *Radiat Oncol.* 2006;1:37.
88. Beyer DC. Salvage brachytherapy after external-beam irradiation for prostate cancer. *Oncology (Williston Park).* 2004;18(2):151–158. ; discussion 158-60, 163-4.
89. Bianchini D, et al. Horizon scanning for novel therapeutics for the treatment of prostate cancer. *Ann Oncol.* 2010;21(Suppl 7):vii43–55.
90. Bilusic M, Heery C, Madan RA. Immunotherapy in prostate cancer: emerging strategies against a formidable foe. *Vaccine.* 2011;29(38):6485–6497.
91. Bittner N, et al. Interstitial brachytherapy should be standard of care for treatment of high-risk prostate cancer. *Oncology (Williston Park).* 2008;22(9):995–1004. ; discussion 1006, 1011-7.
92. Bolla M, Fourmet P, Descotes JL. [Interest of the radiotherapy-hormonotherapy association in the treatment of the high-risk prostate cancer]. *Bull Cancer.* 2008;95(12):1213–1218.
93. Bolton EM, et al. Noncoding RNAs in prostate cancer: the long and the short of it. *Clin Cancer Res.* 2014;20(1):35–43.
94. Bostwick DG, Meiers I. Diagnosis of prostatic carcinoma after therapy. *Arch Pathol Lab Med.* 2007;131(3):360–371.
95. Bott SR, et al. Prostate cancer management: 2. An update on locally advanced and metastatic disease. *Postgrad Med.* 2003;79(937):643–645.
96. Brandes A, et al. Costs of conservative management of early-stage prostate cancer compared to radical prostatectomy—a claims data analysis. *BMC Health Serv Res.* 2016;16(1):664.
97. Brown A, et al. Obesity does not influence prostate intrafractional motion. *J Med Radiat Sci.* 2018;65(1):31–38.
98. Bryant G, Wang L, Mulholland DJ. Overcoming oncogenic mediated tumor immunity in prostate cancer. *Int J Mol Sci.* 2017;18(7).
99. Calais J, Cao M, Nickols NG. The utility of PET/CT in the planning of external radiation therapy for prostate cancer. *J Nucl Med.* 2018;59(4):557–567.
100. Carneiro BA, et al. Emerging subtypes and new treatments for castration-resistant prostate cancer. *Am Soc Clin Oncol Educ Book.* 2020;40:e319–e332.
101. Carthon BC, et al. Therapeutic options for a rising PSA after radical prostatectomy. *Cancer J Urol.* 2013;20(3):6748–6755.
102. Cary C, Odisho AY, Cooperberg MR. Variation in prostate cancer treatment associated with population density of the county of residence. *Prostate Cancer Prostatic Dis.* 2016;19(2):174–179.
103. Castellani D, et al. A structured framework for optimizing high-intensity focused ultrasound ablation treatment in localized prostate cancer. *Investig Clin Urol.* 2019;60(4):312–318.
104. Challapalli A, et al. High dose rate prostate brachytherapy: an overview of the rationale, experience and emerging applications in the treatment of prostate cancer. *Br J Radiol.* 2012;85(Spec No 1):S18–S27. Spec Iss 1.
105. Chambers SK, et al. Living with prostate cancer: randomised controlled trial of a multimodal supportive care intervention for men with prostate cancer. *BMC Cancer.* 2011;11:317.
106. Chen RC, et al. Treatment 'mismatch' in early prostate cancer: do treatment choices take patient quality of life into account? *Cancer.* 2008;112(1):61–68.
107. Chen TC. Prostate cancer and spinal cord compression. *Oncology (Williston Park).* 2001;15(7):841–855. ; discussion 855,859-61.
108. Chen Y, Scher HI. Prostate cancer in 2011: hitting old targets better and identifying new targets. *Nat Rev Clin Oncol.* 2012;9(2):70–72.
109. Cheng ML, Fong L. Beyond sipuleucel-T: immune approaches to treating prostate cancer. *Curr Treat Options Oncol.* 2014;15(1):115–126.
110. Cheng X, et al. The role of radical prostatectomy and definitive external beam radiotherapy in combined treatment for high-risk prostate cancer: a systematic review and meta-analysis. *Asian J Androl.* 2020;22(4):383–389.
111. Chin JL, Touma N. Current status of salvage cryoablation for prostate cancer following radiation failure. *Technol Cancer Res Treat.* 2005;4(2):211–216.
112. Choe KS, Liauw SL. Radiotherapeutic strategies in the management of low-risk prostate cancer. *Sci World J.* 2010;10:1854–1869.
113. Ciezki JP, Klein EA. Brachytherapy or surgery? A composite view. *Oncology (Williston Park).* 2009;23(11):960–964.
114. Clemente S, et al. Monitor unit optimization in RapidArc plans for prostate cancer. *J Appl Clin Med Phys.* 2013;14(3):4114.
115. Conde Moreno AJ, et al. Oligometastases in prostate cancer: restaging stage IV cancers and new radiotherapy options. *Radiat Oncol.* 2014;9:258.
116. Conran CA, Brendler CB, Xu J. Personalized prostate cancer care: from screening to treatment. *Asian J Androl.* 2016;18(4):505–508.
117. Considine B, Petrylak DP. Integrating novel targets and precision medicine into prostate cancer care-Part 1: the non-androgen-targetable pathways in castration-resistant prostate cancer. *Oncology (Williston Park).* 2019;33(3):113–118.
118. Cooperberg MR. Prostate cancer: a new look at prostate cancer treatment complications. *Nat Rev Clin Oncol.* 2014;11(6):304–305.
119. Cooperberg MR, Park S, Carroll PR. Prostate cancer 2004: insights from national disease registries. *Oncology (Williston Park).* 2004;18(10):1239–1247. ; discussion 1248-50, 1256-8.

120. Cordes LM, Gulley JL, Madan RA. The evolving role of immunotherapy in prostate cancer. *Curr Opin Oncol*. 2016;28(3):232–240.
121. Cormie P, et al. Improving sexual health in men with prostate cancer: randomised controlled trial of exercise and psychosexual therapies. *BMC Cancer*. 2014;14:199.
122. Crawford ED, et al. Evolving understanding and categorization of prostate cancer: preventing progression to metastatic castration-resistant prostate cancer: radar IV. *Can J Urol*. 2020;27(5):10352–10362.
123. Czerwińska M, et al. Targeted radionuclide therapy of prostate cancer—from basic research to clinical perspectives. *Molecules*. 2020;25(7).
124. Da Rosa MR, et al. Early experience in MRI-guided therapies of prostate cancer: HIFU, laser and photodynamic treatment. *Cancer Imag*. 2011;11(Spec No A):S3–S8, 1a.
125. Dal Pra A, Cury FL, Souhami L. Radiation therapy and androgen deprivation in the management of high risk prostate cancer. *Int Braz J Urol*. 2011;37(2):161–175. ; discussion 176–9.
126. Devos G, Joniau S. PREDICT Prostate, a useful tool in men with low- and intermediate-risk prostate cancer who are hesitant between conservative management and active treatment. *BMC Med*. 2020;18(1):213.
127. Dillard AJ, et al. Anxiety symptoms prior to a prostate cancer diagnosis: associations with knowledge and openness to treatment. *Br J Health Psychol*. 2017;22(1): 151–168.
128. Dong S, et al. First human trial of high-frequency irreversible electroporation therapy for prostate cancer. *Technol Cancer Res Treat*. 2018;17:1533033818789692.
129. Donin NM, Reiter RE. Why targeting PSMA is a game changer in the management of prostate cancer. *J Nucl Med*. 2018;59(2):177–182.
130. Dorff TB, et al. The evolving role of prostate-specific membrane antigen-based diagnostics and therapeutics in prostate cancer. *Am Soc Clin Oncol Educ Book*. 2019; 39:321–330.
131. Dosoretz AM, et al. Mortality in men with localized prostate cancer treated with brachytherapy with or without neoadjuvant hormone therapy. *Cancer*. 2010; 116(4):837–842.
132. Drake CG. Immunotherapy for metastatic prostate cancer. *Urol Oncol*. 2008;26(4): 438–444.
133. Drake CG. Immunotherapy for prostate cancer: an emerging treatment modality. *Urol Clin North Am*. 2010;37(1):121–129 [Table of Contents].
134. Drake CG, Antonarakis ES. Update: immunological strategies for prostate cancer. *Curr Urol Rep*. 2010;11(3):202–207.
135. Duchesne G. Localised prostate cancer - current treatment options. *Aust Fam Physician*. 2011;40(10):768–771.
136. Efsthathiou JA, Gray PJ, Zietman AL. Proton beam therapy and localised prostate cancer: current status and controversies. *Br J Cancer*. 2013;108(6):1225–1230.
137. Eggner SE, Coleman JA. Focal treatment of prostate cancer with vascular-targeted photodynamic therapy. *Sci World J*. 2008;8:963–973.
138. Elmahdy MS, et al. Robust contour propagation using deep learning and image registration for online adaptive proton therapy of prostate cancer. *Med Phys*. 2019; 46(8):3329–3343.
139. Fahmy O, et al. The role of radical prostatectomy and radiotherapy in treatment of locally advanced prostate cancer: a systematic review and meta-analysis. *Urol Int*. 2017;99(3):249–256.
140. Filson CP, Marks LS, Litwin MS. Expectant management for men with early stage prostate cancer. *CA Cancer J Clin*. 2015;65(4):265–282.
141. Flaig TW, et al. Treatment evolution for metastatic castration-resistant prostate cancer with recent introduction of novel agents: retrospective analysis of real-world data. *Cancer Med*. 2016;5(2):182–191.
142. Focht BC, et al. Effects of a group-mediated exercise and dietary intervention in the treatment of prostate cancer patients undergoing androgen deprivation therapy: results from the IDEAL-P trial. *Ann Behav Med*. 2018;52(5):412–428.
143. Freytag SO, et al. Prostate cancer gene therapy clinical trials. *Mol Ther*. 2007;15(6): 1042–1052.
144. Fu W, et al. Progress of molecular targeted therapies for prostate cancers. *Biochim Biophys Acta*. 2012;1825(2):140–152.
145. Gandellini P, Folini M, Zaffaroni N. Emerging role of microRNAs in prostate cancer: implications for personalized medicine. *Discov Med*. 2010;9(46):212–218.
146. Garcia JA, Dreicer R. Immunotherapy in castration-resistant prostate cancer: integrating sipuleucel-T into our current treatment paradigm. *Oncology (Williston Park)*. 2011;25(3):242–249.
147. Garcia JA, Rini BI. Castration-resistant prostate cancer: many treatments, many options, many challenges ahead. *Cancer*. 2012;118(10):2583–2593.
148. Gavin AT, et al. Patient-reported 'ever had' and 'current' long-term physical symptoms after prostate cancer treatments. *BJU Int*. 2015;116(3):397–406.
149. Gay HA, Michalski JM. Radiation therapy for prostate cancer. *Mo Med*. 2018; 115(2):146–150.
150. Gerritsen WR, Sharma P. Current and emerging treatment options for castration-resistant prostate cancer: a focus on immunotherapy. *J Clin Immunol*. 2012;32(1): 25–35.
151. Ghadjar P, et al. High-dose (80 Gy) intensity-modulated radiation therapy with daily image-guidance as primary treatment for localized prostate cancer. *Strahlenther Onkol*. 2010;186(12):687–692.
152. Gheewala T, Skwor T, Munirathinam G. Photosensitizers in prostate cancer therapy. *Oncotarget*. 2017;8(18):30524–30538.
153. Gkialas IK, Fragkoulis C. Emerging therapies targeting castration-resistant prostate cancer. *J buon*. 2015;20(6):1389–1396.
154. Gomella LG, Petrylak DP, Shayegan B. Current management of advanced and castration resistant prostate cancer. *Can J Urol*. 2014;21(2 Supp 1):1–6.
155. Gómez-Aparicio MA, et al. Extreme hypofractionation with SBRT in localized prostate cancer. *Curr Oncol*. 2021;28(4):2933–2949.
156. Gotoh A, et al. [Gene therapy for prostate cancer]. *Hinyokika Kiyo*. 2002;48(11): 729–732.
157. Gottschalk AR, Roach 3rd M. The use of hormonal therapy with radiotherapy for prostate cancer: analysis of prospective randomised trials. *Br J Cancer*. 2004;90(5): 950–954.
158. Grise P, Thurman S. Urinary incontinence following treatment of localized prostate cancer. *Cancer Control*. 2001;8(6):532–539.
159. Grozescu T, Popa F. Immunotherapy and gene therapy in prostate cancer treatment. *J Med Life*. 2017;10(1):54–55.
160. Gupta A, et al. The role of primary tumor treatment and metastasis-directed therapy in oligometastatic prostate cancer. *Oncology (Williston Park)*. 2019;33(5):187–191.
161. Gustavsen G, et al. Economic burden of illness associated with localized prostate cancer in the United States. *Future Oncol*. 2020;16(1):4265–4277.
162. Halpern JA, et al. Use, complications, and costs of stereotactic body radiotherapy for localized prostate cancer. *Cancer*. 2016;122(16):2496–2504.
163. Haroon UH, et al. Incidence, management, and clinical outcomes of prostate cancer in kidney transplant recipients. *Exp Clin Transplant*. 2019;17(3):298–303.
164. Heysek RV. Modern brachytherapy for treatment of prostate cancer. *Cancer Control*. 2007;14(3):238–243.
165. Hirst AM, et al. Low temperature plasma: a novel focal therapy for localized prostate cancer? *BioMed Res Int*. 2014;2014:878319.
166. Hoang DT, et al. Androgen receptor-dependent and -independent mechanisms driving prostate cancer progression: opportunities for therapeutic targeting from multiple angles. *Oncotarget*. 2017;8(2):3724–3745.
167. Hoffman KE, et al. Prostate cancer-specific mortality and the extent of therapy in healthy elderly men with high-risk prostate cancer. *Cancer*. 2010;116(11): 2590–2595.
168. Hope TA, et al. Targeted PET imaging for prostate-specific membrane antigen in prostate cancer. *Future Oncol*. 2016;12(21):2393–2396.
169. Horwich A. Systemic treatment for prostate cancer. *Ann Oncol*. 2006;17(Suppl 10): x211–x213.
170. Horwich A, et al. Prostate cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2010;21(Suppl 5):v129–v133.
171. Horwitz EM, Hanks GE. External beam radiation therapy for prostate cancer. *CA Cancer J Clin*. 2000;50(6):349–375. quiz 376–9.
172. Hoskin PJ, et al. GEC/ESTRO recommendations on high dose rate afterloading brachytherapy for localised prostate cancer: an update. *Radiother Oncol*. 2013; 107(3):325–332.
173. Hu J, et al. Preoperative therapy for localized prostate cancer: a comprehensive overview. *Maturitas*. 2013;74(1):3–9.
174. Hurwitz LM, et al. Longitudinal regret after treatment for low- and intermediate-risk prostate cancer. *Cancer*. 2017;123(21):4252–4258.
175. Hurwitz MD, et al. Hyperthermia combined with radiation for the treatment of locally advanced prostate cancer: long-term results from Dana-Farber Cancer Institute study 94-153. *Cancer*. 2011;117(3):510–516.
176. Hussain A, Dawson N. Management of advanced/metastatic prostate cancer: 2000 update. *Oncology (Williston Park)*. 2000;14(12):1677–1688. ; discussion 1688, 1691–4.
177. Jang JW, et al. Long-term quality of life after definitive treatment for prostate cancer: patient-reported outcomes in the second posttreatment decade. *Cancer Med*. 2017;6(7):1827–1836.
178. Jang TL, et al. Low risk prostate cancer in men under age 65: the case for definitive treatment. *Urol Oncol*. 2007;25(6):510–514.
179. Jindal V. Immunotherapy: a glimmer of hope for metastatic prostate cancer. *Chin Clin Oncol*. 2018;7(6):61.
180. Johansen TE. [Cryo-therapy for prostate cancer]. *Tidsskr Nor Laegeforen*. 2005; 125(12):1661–1663.
181. Johansen TE. [Radical therapy for prostate cancer in Norway]. *Tidsskr Nor Laegeforen*. 2005;125(12):1658–1660.
182. Jorgo K, et al. [Stereotactic body radiation therapy with CyberKnife accelerator for low- and intermediate risk prostate cancer]. *Magy Onkol*. 2019;63(1):52–59.
183. Joseph KJ, et al. Analysis of health related quality of life (HRQoL) of patients with clinically localized prostate cancer, one year after treatment with external beam radiotherapy (EBRT) alone versus EBRT and high dose rate brachytherapy (HDRBT). *Radiat Oncol*. 2008;3:20.
184. Jung JH, et al. Primary cryotherapy for localised or locally advanced prostate cancer. *Cochrane Database Syst Rev*. 2018;5(5):Cd005010.
185. Kang HW, et al. Current status of radical prostatectomy for high-risk prostate cancer. *Korean J Urol*. 2014;55(10):629–635.
186. Karavitakis M, et al. Focal therapy for prostate cancer: opportunities and uncertainties. *Discov Med*. 2011;12(64):245–255.
187. Katz AJ. CyberKnife radiosurgery for prostate cancer. *Technol Cancer Res Treat*. 2010;9(5):463–472.
188. Porfyrus O, Kalomoiris P. Current role of immunotherapy for the treatment of prostate cancer. *J buon*. 2013;18(4):809–817.
189. Kilcoyne A, Harisinghani MG, Mahmood U. Prostate cancer imaging and therapy: potential role of nanoparticles. *J Nucl Med*. 2016;57(Suppl 3):105S–110S.
190. Kim J, Park JS, Ham WS. The role of metastasis-directed therapy and local therapy of the primary tumor in the management of oligometastatic prostate cancer. *Investig Clin Urol*. 2017;58(5):307–316.
191. Kim JH, Lee HJ, Song YS. Stem cell based gene therapy in prostate cancer. *BioMed Res Int*. 2014;2014:549136.
192. Laccetti AL, Subudhi SK. Immunotherapy for metastatic prostate cancer: immuno-club or the tip of the iceberg? *Curr Opin Urol*. 2017;27(6):566–571.
193. Klein EA. Prostate cancer: current concepts in diagnosis and treatment. *Cleve Clin J Med*. 1992;59(4):383–389.

194. Kohli M, Tindall DJ. New developments in the medical management of prostate cancer. *Mayo Clin Proc.* 2010;85(1):77–86.
195. Koukourakis MI, Touloupidis S. External beam radiotherapy for prostate cancer: current position and trends. *Anticancer Res.* 2006;26(1b):485–494.
196. Lee IY, et al. Dihydroisotanshinone I combined with radiation inhibits the migration ability of prostate cancer cells through DNA damage and CCL2 pathway. *BMC Pharmacol Toxicol.* 2018;19(1):5.
197. Kvåle R, et al. [Curative treatment of prostatic cancer in Norway in 1998 and 2001]. *Tidsskr Nor Laegeforen.* 2006;126(7):912–916.
198. Lei JH, et al. Systematic review and meta-analysis of the survival outcomes of first-line treatment options in high-risk prostate cancer. *Sci Rep.* 2015;5:7713.
199. Leão R, et al. Cancer stem cells in prostate cancer: implications for targeted therapy. *Urol Int.* 2017;99(2):125–136.
200. Lehto US, et al. Patients' perceptions of the negative effects following different prostate cancer treatments and the impact on psychological well-being: a nationwide survey. *Br J Cancer.* 2017;116(7):864–873.
201. Kubes J, et al. Treatment of high risk prostate cancer with combined radiotherapy and hormonal treatment- results and identification of factors influencing outcome. *J buon.* 2013;18(3):669–674.
202. Lewis DD, Cropp CD. The impact of African ancestry on prostate cancer disparities in the era of precision medicine. *Genes.* 2020;11(12).
203. Kim W, Ryan CJ. Quo vadis: advanced prostate cancer-clinical care and clinical research in the era of multiple androgen receptor-directed therapies. *Cancer.* 2015;121(3):361–371.
204. Linares-Espinós E, et al. New technologies and techniques for prostate cancer focal therapy. *Minerva Urol Nefrol.* 2018;70(3):252–263.
205. Ketola K, et al. Targeting prostate cancer subtype 1 by Forkhead box M1 pathway inhibition. *Clin Cancer Res.* 2017;23(22):6923–6933.
206. Liu JJ, Zhang J. Sequencing systemic therapies in metastatic castration-resistant prostate cancer. *Cancer Control.* 2013;20(3):181–187.
207. Lo ST, et al. Dendrimer nanoscaffolds for potential theranostics of prostate cancer with a focus on radiochemistry. *Mol Pharm.* 2013;10(3):793–812.
208. Loda M, Kaelin Jr WG. Prostate cancer: beta control your hormones. *Cancer Cell.* 2010;17(4):311–312.
209. Lopez W, et al. Ultrasound therapy, chemotherapy and their combination for prostate cancer. *Technol Cancer Res Treat.* 2021;20:15330338211011965.
210. Lorient Y, Massard C, Fizazi K. Recent developments in treatments targeting castration-resistant prostate cancer bone metastases. *Ann Oncol.* 2012;23(5):1085–1094.
211. Lou DY, Fong L. Neoadjuvant therapy for localized prostate cancer: examining mechanism of action and efficacy within the tumor. *Urol Oncol.* 2016;34(4):182–192.
212. Low JY, et al. Effective targeting of RNA polymerase I in treatment-resistant prostate cancer. *Prostate.* 2019;79(16):1837–1851.
213. Lu W, et al. Gold nano-porcnon-based targeted diagnosis, nanotherapy treatment, and in situ monitoring of photothermal therapy response of prostate cancer cells using surface-enhanced Raman spectroscopy. *J Am Chem Soc.* 2010;132(51):18103–18114.
214. Luk SU, et al. Chemopreventive effect of PSP through targeting of prostate cancer stem cell-like population. *PLoS One.* 2011;6(5):e19804.
215. Lumen N, et al. Developments in external beam radiotherapy for prostate cancer. *Urology.* 2013;82(1):5–10.
216. Lu-Yao GL, et al. Primary radiotherapy vs conservative management for localized prostate cancer—a population-based study. *Prostate Cancer Prostatic Dis.* 2015;18(4):317–324.
217. Ma L, et al. Optimized intensity-modulated arc therapy for prostate cancer treatment. *Int J Cancer.* 2001;96(6):379–384.
218. Manabe Y, et al. Stereotactic body radiotherapy using a hydrogel spacer for localized prostate cancer: a dosimetric comparison between tomotherapy with the newly-developed tumor-tracking system and cyberknife. *J Appl Clin Med Phys.* 2021;22(10):66–72.
219. Mangoni M, et al. Hypofractionation in prostate cancer: radiobiological basis and clinical appliance. *BioMed Res Int.* 2014;2014:781340.
220. Mano R, Eastham J, Yossepowitch O. The very-high-risk prostate cancer: a contemporary update. *Prostate Cancer Prostatic Dis.* 2016;19(4):340–348.
221. Marra G, et al. Multimodal treatment in focal therapy for localized prostate cancer using concomitant short-term androgen deprivation therapy: the ENHANCE prospective pilot study. *Minerva Urol Nefrol.* 2019;71(5):544–548.
222. Marzo Castillejo M, et al. [Updating in prevention and treatment of prostate cancer]. *Aten Primaria.* 2002;30(1):57–63.
223. Massard C, Fizazi K. Targeting continued androgen receptor signaling in prostate cancer. *Clin Cancer Res.* 2011;17(12):3876–3883.
224. McLaughlin PW, Narayana V. Progress in low dose rate brachytherapy for prostate cancer. *Semin Radiat Oncol.* 2020;30(1):39–48.
225. Mitin T, et al. Management of lymph node-positive prostate cancer: the role of surgery and radiation therapy. *Oncology (Williston Park).* 2013;27(7):647–655.
226. Miyahira AK, et al. Beyond the androgen receptor II: new approaches to understanding and treating metastatic prostate cancer; Report from the 2017 Coffey-Holden Prostate Cancer Academy Meeting. *Prostate.* 2017;77(15):1478–1488.
227. Murphy GP. Prostate cancer. *CA Cancer J Clin.* 1974;24(5):282–288.
228. Nakamura T, et al. [Symptoms of prostate cancer that required treatment in the terminal stage for two years]. *Hinyokika Kyo.* 2010;56(1):11–15.
229. Munoz J, Wheler JJ, Kurzrock R. Androgen receptors beyond prostate cancer: an old marker as a new target. *Oncotarget.* 2015;6(2):592–603.
230. Murphy C, et al. A novel system for estimating residual disease and pathologic response to neoadjuvant treatment of prostate cancer. *Prostate.* 2016;76(14):1285–1292.
231. Simons JW. Prostate cancer immunotherapy: beyond immunity to curability. *Cancer Immunol Res.* 2014;2(11):1034–1043.
232. Sita-Lumsden A, et al. Circulating microRNAs as potential new biomarkers for prostate cancer. *Br J Cancer.* 2013;108(10):1925–1930.
233. Skolarus TA, et al. Optimizing veteran-centered prostate cancer survivorship care: study protocol for a randomized controlled trial. *Trials.* 2017;18(1):181.
234. Su D, Jang TL. Using large institutional or national databases to evaluate prostate cancer outcomes and patterns of care: possibilities and limitations. *Sci World J.* 2011;11:147–160.
235. Thoreson GR, et al. Emerging therapies in castration resistant prostate cancer. *Can J Urol.* 2014;21(2 Supp 1):98–105.
236. Morgans AK, Stockler MR. Patient-reported outcomes in metastatic castration-sensitive prostate cancer in the adjuvant setting. *Eur Urol Focus.* 2019;5(2):144–146.
237. Tran PT, et al. Tissue biomarkers for prostate cancer radiation therapy. *Curr Mol Med.* 2012;12(6):772–787.
238. Tse BW, et al. From bench to bedside: immunotherapy for prostate cancer. *BioMed Res Int.* 2014;2014:981434.
239. Turkbey B, Pinto PA, Choyke PL. Imaging techniques for prostate cancer: implications for focal therapy. *Nat Rev Urol.* 2009;6(4):191–203.
240. Tyson MD, Penson DF, Resnick MJ. The comparative oncologic effectiveness of available management strategies for clinically localized prostate cancer. *Urol Oncol.* 2017;35(2):51–58.
241. Volk RJ, et al. It's not like you just had a heart attack': decision-making about active surveillance by men with localized prostate cancer. *Psycho Oncol.* 2014;23(4):467–472.
242. Wade CA, Kyprianou N. Profiling prostate cancer therapeutic resistance. *Int J Mol Sci.* 2018;19(3).
243. Mohebtash M, et al. Therapeutic prostate cancer vaccines: a review of the latest developments. *Curr Opin Invest Drugs.* 2008;9(12):1296–1301.
244. Slovin SF. Emerging role of immunotherapy in the management of prostate cancer. *Oncology (Williston Park).* 2007;21(3):326–333. ; discussion 334, 338, 346–8.
245. Smolle MA, et al. Current insights into long non-coding RNAs (LncRNAs) in prostate cancer. *Int J Mol Sci.* 2017;18(2).
246. Snyder CF, et al. How does initial treatment choice affect short-term and long-term costs for clinically localized prostate cancer? *Cancer.* 2010;116(23):5391–5399.
247. Soloway M, Roach 3rd M. Prostate cancer progression after therapy of primary curative intent: a review of data from prostate-specific antigen era. *Cancer.* 2005;104(11):2310–2322.
248. Song J, et al. Identification of immune-based prostate cancer subtypes using mRNA expression. *Biosci Rep.* 2021;41(1).
249. Spratt DE, et al. Treating the patient and not just the cancer: therapeutic burden in prostate cancer. *Prostate Cancer Prostatic Dis.* 2021;24(3):647–661.
250. Stokes WA, et al. Racial differences in time from prostate cancer diagnosis to treatment initiation: a population-based study. *Cancer.* 2013;119(13):2486–2493.
251. Tanino T, Uchida N. [7. Radiation therapy for prostate cancer]. *Nippon Hoshasen Gijutsu Gakkai Zasshi.* 2018;74(1):84–93.
252. Tarassoff CP, Arlen PM, Gulley JL. Therapeutic vaccines for prostate cancer. *Oncol.* 2006;11(5):451–462.
253. Tian Z, et al. Comparison of radical prostatectomy versus conservative treatment in localized prostate cancer: systematic review and meta-analysis. *J buon.* 2019;24(1):239–248.
254. Zapotoczna A, et al. Current role and future perspectives of magnetic resonance spectroscopy in radiation oncology for prostate cancer. *Neoplasia.* 2007;9(6):455–463.
255. Zhang J, et al. Inhibition of GLS suppresses proliferation and promotes apoptosis in prostate cancer. *Biosci Rep.* 2019;39(6).
256. Zhang KX, Jia W, Rennie PS. Bioengineered viral vectors for targeting and killing prostate cancer cells. *Bioeng Bugs.* 2010;1(2):92–96.
257. Zhang W, et al. Role of the DNA damage response in prostate cancer formation, progression and treatment. *Prostate Cancer Prostatic Dis.* 2020;23(1):24–37.
258. Zhu W, et al. Treatment of castration-resistant prostate cancer: updates on therapeutics targeting the androgen receptor signaling pathway. *Am J Therapeut.* 2010;17(2):176–181.
259. Nguyen CT, Zelefsky MJ, Kattan MW. The current state of brachytherapy nomograms for patients with clinically localized prostate cancer. *Cancer.* 2009;115(13 Suppl):3121–3127.
260. Novaes P, Mottas RT, Lundgren M. Treatment of prostate cancer with intensity modulated radiation therapy (IMRT). *Rev Assoc Med Bras.* 2015;61(1):8–16, 1992.
261. Udager AM, Tomlins SA. Molecular biomarkers in the clinical management of prostate cancer. *Cold Spring Harb Perspect Med.* 2018;8(11).
262. Valerio M, et al. The role of focal therapy in the management of localised prostate cancer: a systematic review. *Eur Urol.* 2014;66(4):732–751.
263. van der Poel HG, et al. Focal therapy in primary localised prostate cancer: the European association of urology position in 2018. *Eur Urol.* 2018;74(1):84–91.
264. Varnava M, et al. Dosimetric comparison between volumetric modulated arc therapy planning techniques for prostate cancer in the presence of intrafractional organ deformation. *J Radiat Res.* 2021;62(2):309–318.
265. Vellyk JE, Ricke WA. Development and prevalence of castration-resistant prostate cancer subtypes. *Neoplasia.* 2020;22(11):566–575.
266. Vitkin N, et al. The tumor immune contexture of prostate cancer. *Front Immunol.* 2019;10:603.



267. Walia G, et al. The 19th annual Prostate Cancer Foundation scientific retreat. *Cancer Res.* 2013;73(16):4988–4991.
268. Wang Y, et al. Hypofractionated proton therapy for prostate cancer: dose delivery uncertainty due to interfractional motion. *Med Phys.* 2013;40(7):071714.
269. Want MY, et al. Inhibition of WHSC1 allows for reprogramming of the immune compartment in prostate cancer. *Int J Mol Sci.* 2021;22(16).
270. Wenger H, et al. Laser ablation as focal therapy for prostate cancer. *Curr Opin Urol.* 2014;24(3):236–240.
271. White M, Verhoef M. Cancer as part of the journey: the role of spirituality in the decision to decline conventional prostate cancer treatment and to use complementary and alternative medicine. *Integr Cancer Ther.* 2006;5(2):117–122.
272. Wilson LS, et al. Cumulative cost pattern comparison of prostate cancer treatments. *Cancer.* 2007;109(3):518–527.
273. Wilt TJ, Brawer MK. The prostate cancer intervention versus observation trial (PIVOT). *Oncology (Williston Park).* 1997;11(8):1133–1139. ; discussion 1139–40, 1143.
274. Wilt TJ, et al. Systematic review: comparative effectiveness and harms of treatments for clinically localized prostate cancer. *Ann Intern Med.* 2008;148(6):435–448.
275. Wolff RF, et al. A systematic review of randomised controlled trials of radiotherapy for localised prostate cancer. *Eur J Cancer.* 2015;51(16):2345–2367.
276. Wozney JL, Antonarakis ES. Growth factor and signaling pathways and their relevance to prostate cancer therapeutics. *Cancer Metastasis Rev.* 2014;33(2–3): 581–594.
277. Yamada Y, et al. Salvage brachytherapy for locally recurrent prostate cancer after external beam radiotherapy. *Asian J Androl.* 2015;17(6):899–903.
278. Yamoah K, et al. Early results of prostate cancer radiation therapy: an analysis with emphasis on research strategies to improve treatment delivery and outcomes. *BMC Cancer.* 2013;13:23.
279. Yartsev S, Bauman G. Target margins in radiotherapy of prostate cancer. *Br J Radiol.* 2016;89(1067):20160312.
280. Yee DS, et al. Impact of previous radiotherapy for prostate cancer on clinical outcomes of patients with bladder cancer. *J Urol.* 2010;183(5):1751–1756.
281. Yeku O, Slovin SF. Immune therapy for prostate cancer. *Cancer J.* 2016;22(5): 334–341.
282. Oh WK. High-risk localized prostate cancer: integrating chemotherapy. *Oncol.* 2005; 10(Suppl 2):18–22.
283. O'Hanlon Brown C, Waxman J. Current management of prostate cancer: dilemmas and trials. *Br J Radiol.* 2012;85(Spec No 1):S28–S40. Spec Iss 1.
284. Okada K, et al. [Treatment of advanced prostate cancer]. *Hinyokika Kyo.* 1994; 40(10):945–950.
285. O'Keefe DS, et al. A perspective on the evolving story of PSMA biology, PSMA-based imaging, and endoradiotherapeutic strategies. *J Nucl Med.* 2018;59(7):1007–1013.
286. Olson B, Patnaik A. Utilizing precision medicine to modulate the prostate tumor microenvironment and enhance immunotherapy. *Urol Oncol.* 2019;37(8):535–542.
287. Palacios DA, Miyake M, Rosser CJ. Radioisotization in prostate cancer: mechanisms and targets. *BMC Urol.* 2013;13:4.
288. Ong WM, et al. Prostate specific membrane antigen: the role in salvage lymph node dissection and radio-ligand therapy. *Minerva Urol Nefrol.* 2018;70(5):450–461.
289. Oottamasathien S, Crawford ED. Recent advances in hormonal therapy for advanced prostate cancer. *Oncology (Williston Park).* 2003;17(8):1047–1052. ; discussion 1054–8.
290. Ory J, et al. Testosterone therapy in patients with treated and untreated prostate cancer: impact on oncologic outcomes. *J Urol.* 2016;196(4):1082–1089.
291. Osakabe H, et al. [Undifferentiated prostate cancer treated with radiation therapy]. *Hinyokika Kyo.* 2016;62(11):599–604.
292. Paller CJ, et al. Management of patients with biochemical recurrence after local therapy for prostate cancer. *Hematol Oncol Clin N Am.* 2013;27(6):1205, 19, viii.
293. Palmboos PL, Hussain MH. Targeting PARP in prostate cancer: novelty, pitfalls, and promise. *Oncology (Williston Park).* 2016;30(5):377–385.
294. Payne H. Management of locally advanced prostate cancer. *Asian J Androl.* 2009; 11(1):81–87.
295. Perloth DJ, et al. An economic analysis of conservative management versus active treatment for men with localized prostate cancer. *J Natl Cancer Inst Monogr.* 2012; 2012(45):250–257.
296. Pezaro CJ, Marciscano AE, Madan RA. The winds of change: emerging therapeutics in prostate cancer. *Am Soc Clin Oncol Educ Book.* 2018;38:382–390.
297. Phillips R, Ost P, Tran PT. What role does stereotactic ablative radiotherapy have in advanced castrate-resistant prostate cancer? *Future Oncol.* 2017;13(24):2121–2124.
298. Pickles T. What's a man to do? Treatment options for localized prostate cancer. *Can Fam Physician.* 2004;50:65–72.
299. Pienta KJ, et al. Beyond the androgen receptor: new approaches to treating metastatic prostate cancer. Report of the 2013 Prostate Cancer Meeting. *Prostate.* 2014;74(3):314–320.
300. Pisters LL. Treatment failure after primary and salvage therapy for prostate cancer. *Cancer.* 2008;112(2):225–227.
301. Potters L. Permanent prostate brachytherapy: lessons learned, lessons to learn. *Oncology (Williston Park).* 2000;14(7):981–991. ; discussion 991–2, 997–9.
302. Ragde H, et al. Modern prostate brachytherapy. *CA Cancer J Clin.* 2000;50(6): 380–393.
303. Potugari BR, Engel JM, Onitilo AA. Metastatic prostate cancer in a RAD51C mutation carrier. *Clin Med Res.* 2018;16(3–4):69–72.
304. Printz C. Combined therapies improve survival in aggressive prostate cancer. *Cancer.* 2018;124(12):2470.
305. Prada PJ, et al. Low-dose-rate brachytherapy for patients with transurethral resection before implantation in prostate cancer. Longterm results. *Int Braz J Urol.* 2016;42(1):47–52.
306. Printz C. Delaying radiation therapy for prostate cancer unlikely to affect survival during coronavirus disease 2019. *Cancer.* 2021;127(2):171.
307. Raghavan D. Prostate cancer: too much dogma, not enough data. *Cleve Clin J Med.* 2008;75(1):33–34.
308. Raldow A, et al. Salvage external beam radiotherapy for prostate cancer after radical prostatectomy: current status and controversy. *Oncology (Williston Park).* 2010;24(8):692–700, 702.
309. Pronzato P, Rondini M. Hormonotherapy of advanced prostate cancer. *Ann Oncol.* 2005;16(Suppl 4):iv80–84.
310. Ramalingam S, et al. What should we tell patients about physical activity after a prostate cancer diagnosis? *Oncology (Williston Park).* 2015;29(9):680–685, 687–685.
311. Rana S, et al. Dosimetric impact of Acuros XB dose calculation algorithm in prostate cancer treatment using RapidArc. *J Cancer Res Ther.* 2013;9(3):430–435.
312. Sankineni S, et al. Image-guided focal therapy for prostate cancer. *Diagn Interv Radiol.* 2014;20(6):492–497.
313. Rao A, et al. Oligometastatic prostate cancer: a shrinking subset or an opportunity for cure? *Am Soc Clin Oncol Educ Book.* 2019;39:309–320.
314. Santer FR, Erb HH, McNeill RV. Therapy escape mechanisms in the malignant prostate. *Semin Cancer Biol.* 2015;35:133–144.
315. Rapiiti E, et al. Impact of socioeconomic status on prostate cancer diagnosis, treatment, and prognosis. *Cancer.* 2009;115(23):5556–5565.
316. Rashid MH, Chaudhary UB. Intermittent androgen deprivation therapy for prostate cancer. *Oncol.* 2004;9(3):295–301.
317. Sarkis M, Ghanem E, Rahme K. Jumping on the bandwagon: a review on the versatile applications of gold nanostructures in prostate cancer. *Int J Mol Sci.* 2019; 20(4).
318. Rathi N, et al. Evolving role of immunotherapy in metastatic castration refractory prostate cancer. *Drugs.* 2021;81(2):191–206.
319. Ray ME, et al. Potential surrogate endpoints for prostate cancer survival: analysis of a phase III randomized trial. *J Natl Cancer Inst.* 2009;101(4):228–236.
320. Reis RBD, et al. Prostate cancer in Latin America: challenges and recommendations. *Cancer Control.* 2020;27(1):1073274820915720.
321. Ren SC, Chen R, Sun YH. Prostate cancer research in China. *Asian J Androl.* 2013; 15(3):350–353.
322. Sartor O, et al. Novel therapeutic strategies for metastatic prostate cancer in the post-docetaxel setting. *Oncol.* 2011;16(11):1487–1497.
323. Rewcastle JC. High intensity focused ultrasound for prostate cancer: a review of the scientific foundation, technology and clinical outcomes. *Technol Cancer Res Treat.* 2006;5(6):619–625.
324. Scardino PT. The Gordon Wilson Lecture. Natural history and treatment of early stage prostate cancer. *Trans Am Clin Climatol Assoc.* 2000;111:201–241.
325. Scher HI. Building on prostate cancer working group 2 to change the paradigm from palliation to cure. *Am Soc Clin Oncol Educ Book.* 2014:e204–e212.
326. Schneider JA, Logan SK. Revisiting the role of Wnt/ $\beta$ -catenin signaling in prostate cancer. *Mol Cell Endocrinol.* 2018;462(Pt A):3–8.
327. Ruah J. [Prostate cancer]. *Acta Med Port.* 1999;12(1–3):91–94.
328. Rutznar S, et al. Noncurated data lead to misinterpretation of treatment outcomes in patients with prostate cancer after salvage or palliative radiotherapy. *JCO Clin Cancer Inform.* 2019;3:1–11.
329. Schumacher O, et al. Exercise modulation of tumour perfusion and hypoxia to improve radiotherapy response in prostate cancer. *Prostate Cancer Prostatic Dis.* 2021;24(1):1–14.
330. Schweizer MT, Drake CG. Immunotherapy for prostate cancer: recent developments and future challenges. *Cancer Metastasis Rev.* 2014;33(2–3):641–655.
331. Sedelaar JP, Schalken JA. The need for a personalized approach for prostate cancer management. *BMC Med.* 2015;13:109.
332. Selley S, et al. Diagnosis, management and screening of early localized prostate cancer. *Health Technol Assess.* 1997;1(2):1–96. i.
333. Seppälä J, et al. Dosimetric comparison and evaluation of 4 stereotactic body radiotherapy techniques for the treatment of prostate cancer. *Technol Cancer Res Treat.* 2017;16(2):238–245.
334. Serra M, et al. Dosimetric comparison among cyberknife, helical tomotherapy and VMAT for hypofractionated treatment in localized prostate cancer. *Medicine (Baltim).* 2020;99(50):e23574.
335. Shah K, Bradbury NA. Lemur Tyrosine Kinase 2, a novel target in prostate cancer therapy. *Oncotarget.* 2015;6(16):14233–14246.
336. Shah MR, et al. Direct intra-tumoral injection of zinc-acetate halts tumor growth in a xenograft model of prostate cancer. *J Exp Clin Cancer Res.* 2009;28(1):84.
337. Sherer BA, et al. Prostate cancer in renal transplant recipients. *Int Braz J Urol.* 2017; 43(6):1021–1032.
338. Shirakawa T, Fujisawa M, Gotoh A. Gene therapy in prostate cancer: past, present and future. *Front Biosci.* 2008;13:2115–2119.
339. Shtivelman E, Beer TM, Evans CP. Molecular pathways and targets in prostate cancer. *Oncotarget.* 2014;5(17):7217–7259.
340. Silvestri I, et al. Beyond the immune suppression: the immunotherapy in prostate cancer. *BioMed Res Int.* 2015;2015:794968.
341. Simmons MN, Berglund RK, Jones JS. A practical guide to prostate cancer diagnosis and management. *Cleve Clin J Med.* 2011;78(5):321–331.