

Optimization of cholesterol oxidase production and 16S rRNA partial sequence of *Bacillus cereus* strain KAVK4 isolated from butter

Amutha Kuppusamy*, Kokila Vasantha Kumar

Department of Biotechnology, Vels University, Chennai, Tamil Nadu, India.

ARTICLE INFO

Article history:

Received on: 15/03/2016

Revised on: 20/04/2016

Accepted on: 30/05/2016

Available online: 28/07/2016

Key words:

Bacillus cereus strain KAVK4, Sequencing, 16S rRNA gene, Cholesterol oxidase, Optimization.

ABSTRACT

The aim of the present research work was to optimize cholesterol oxidase production and partial sequencing of 16S rRNA gene region of *Bacillus cereus* strain KAVK4 by PCR. *Bacillus cereus* is a gram positive bacterium isolated from butter, Tamil Nadu, India and was cultured in a nutrient medium containing cholesterol at 37°C for 24 hours. Partial sequence of 16S rRNA gene was amplified by PCR and sequenced, the sequence data was submitted to NCBI. The production of cholesterol oxidase of *Bacillus cereus* strain KAVK4 was analyzed by qualitative methods such as Colony staining method & Cholesterol Oxidase indicator plate method and quantitative method. The maximum production of cholesterol oxidase was checked in media with different carbon source, nitrogen source, and metal ions source. The different physical parameters like pH, temperature and time were checked. *Bacillus cereus* strain KAVK4 partial sequence of 16S rRNA gene region resulted 1366bp. The sequence data was submitted to NCBI and the accession number is KP792775. *Bacillus cereus* strain KAVK4 was optimized at media containing Fructose, Beef Extract, Ammonium Nitrate and Magnesium sulphate at pH 7.5 at Room temperature and incubation period was about 32 hrs. *Bacillus cereus* strain KAVK4 was found to produce maximum level of cholesterol oxidase (1.67 U/ml). The result obtained in this work was clearly indicated that *Bacillus cereus* strain KAVK4 was capable of producing maximum level of cholesterol oxidase at pH 7.5 and incubation was 32 hrs.

INTRODUCTION

The microbial degradation of cholesterol into 4-cholesten-3-one by the cholesterol oxidase. The oxidation of cholesterol to 4-cholesten-3-one and the reduction of oxygen to hydrogen peroxide which catalyzes by the cholesterol oxidase (Smith and Brooks, 1976 and Sakodinskaya, 2000). It has been widely applied in medical field for the determination of cholesterol in blood serum and food. It acts as a starting material for the chemical synthesis of pharmaceutical steroid (Watanabe *et al.*, 1986). It can also be employed for degrading dietary cholesterol to improve human health associated with cardiovascular disease (Kaunitz *et al.*, 1978). Many microorganisms have an ability to decompose cholesterol that

has been due to the enzyme source in steroid biotransformations, this was achieved by cholesterol oxidase (Constantinides, 1980). Cholesterol oxidase can be produced by bacteria of the genera *Arthrobacter*, *Brevibacterium*, *Pseudomonas*, *Nocardia*, *Rhodococcus*, *Streptomyces*, *Corynebacterium* and *Shizophyllum* (Yazdi *et al.*, 2008). This can be produced in three forms such as intracellular, extracellular and membrane bounded. Cholesterol oxidase are widely applied in various fields, hence screening and isolate of cholesterol oxidase producing bacterial strains are of great interest (Yazdi *et al.*, 2001a). Due to low production of Cholesterol oxidase many researchers may attempt to improve its yield by the modification of the growth medium (Lata Kumari *et al.*, 2012). In the present study, *Bacillus cereus* strain KAVK4 was isolated from Butter and identified at molecular level by sequencing 16S rRNA gene region. Then, the optimization of cholesterol oxidase production by *Bacillus cereus* strain KAVK4 using different media, carbon source, nitrogen source, metal ion source, pH, Temperature and Time were performed in order to increase the cholesterol oxidase production.

* Corresponding Author

Dr. K. Amutha, Associate Professor Department of Biotechnology, Vels University, Pallavaram, Chennai Tamil Nadu, India
E Mail: amutharavi40@gmail.com

MATERIALS AND METHODS

Isolation and identification of *Bacillus cereus* strain KAVK4

The Butter sample was collected from Chennai, Tamil Nadu, India. The sample was serially diluted using 9ml sterile distilled water. The bacterial species was cultured by cholesterol containing nutrient medium at 37°C for 24 hours. *Bacillus cereus* was identified based on physical, morphological and biochemical characterization (Bergey's Manual of determinative bacteriology, Williams and Wilkins 1994).

Molecular identification of *Bacillus cereus*

DNA was extracted from *Bacillus cereus* based on the method described by Pitcher *et al.* (2008). After extraction the DNA sample was run on 1% agarose gel at a constant voltage of 100V. The gel was examined on UV transilluminator.

PCR amplification and sequencing of 16S rRNA

16S rRNA gene region of *Bacillus cereus* was amplified. For setting up PCR, the following reaction mixtures were added into the PCR tube (Applied Biosystems, Acme Progen Biotech (India) Pvt.Ltd, Salem, Tamilnadu, India).

Template- 5µl
518Forward primer-1 µl
800Reverse primer-1 µl
Assay buffer- 6 µl
Taq DNA polymerase- 2 µl
dNTP mix- 5 µl

Using the following reaction conditions, the amplification was carried out in a thermal cycler for 40 cycles. Denaturation of DNA at 94°C for 1 minute, primer annealing at 56 °C for 30 seconds and primer extension at 72 °C for 1 minute. 1.5% agarose gel was casted and the amplified PCR product was mixed with 5 µl of gel loading buffer. DNA ladder of 1kb (HIMEDIA, Mumbai, Maharashtra, India) as a molecular marker was loaded along with samples. The gel was run and examined on UV transilluminator to visualize the bands. PCR products were purified by using the PCR Klenzol™ (Genei, Bangalore, India) and it was sequenced with an ABI Prism 3700 DNA Analyzer (Acme Progen Biotech (India) Pvt. Ltd., Salem, Tamilnadu, India). The nucleotide sequence 16S rRNA gene region data of *Bacillus cereus* was submitted to NCBI database.

Detection of Cholesterol oxidase production by qualitative and quantitative analysis

Qualitative analysis of Cholesterol oxidase

Colony staining method

Colony staining method is to confirm the organism which producing Cholesterol oxidase. The filter disc were immersed into the solution containing 0.5% Cholesterol, 1.7% 4-aminoantipyrin, 6% Phenol and 3000 U/L Horseradish peroxidase (HRP) in 100mM Potassium buffer phosphate (pH 7.0). To the overnight grown bacterial colonies the immersed disc was placed on to the petriplate with *Bacillus cereus* strain KAVK4 and incubates at

room temperature for 24 hrs. Development of red colour due to the formation of quinoneimine dye was confirmed the production of cholesterol oxidase (Lashkarian *et al.*, 2010).

Cholesterol oxidase indicator plates

Bacillus cereus strain KAVK4 which producing Cholesterol oxidase were selected on Indicator plates containing 1.0g/L Cholesterol, 1.0g/L Triton X-100, 0.1g/L o-dianisidine and 1000 U/L horseradish peroxidase of LB agar medium. The overnight bacteria were cultured on these plates and incubate at 30°C. The production of cholesterol oxidase which leads to the conversion of H₂O₂ from the bacterial cells and the reagent in the medium react with H₂O₂ to form azo compound which turns brown color (Lashkarian *et al.*, 2010).

Quantitative analysis of cholesterol oxidase

Cholesterol oxidase activity was analyzed by measuring H₂O₂ generation (Allain *et al.*, 1974). Briefly, the assay procedure is as follows to the 50µl crude enzyme were incubated with 3ml of solution A (4-amino-antipyrine, 1mmol/L; Phenol, 6mmol/L; Sodium azide, 0.2 g/L; Peroxidase, 5000U/L; Potassium phosphate buffer, 25mmol/L PH, 7.5) and 150 µl of solution B(Cholesterol, 8.26mg/ml; Triton X-100, 4.26%; isopropanol for solvent) for 5 min at 37°C, then boiled and was measured by spectrophotometry at 500nm. The enzyme activity was calculated as:

$$\text{Enzyme activity (U/ml)} = 0.1315A_{500} \times 3.2 \times 20 \div 5 = 1.6832 \times A_{500}$$

Optimization of fermentation medium components for the production of cholesterol oxidase

Effect of different media for the production of cholesterol oxidase

Production of Cholesterol oxidase was investigated by modifying the different media. The media such as Media 1(Ahmad and Johri 1992), Media 2(Modification of Arima, 1969), Media 3(Varma and Nene, 2003), Media 4 (Wu *et al.*, 2015) and Media 5 (Ming-Tsung Lee *et al.*, 1997) were taken. The amount of enzyme produced by *Bacillus cereus* strain KAVK4 at 12, 24, 32, 48, 64, 80 hrs of incubation time was determined by the enzymatic assay. The Media 1 showing maximum production of Cholesterol oxidase was selected for further optimization study. For the production of cholesterol oxidase, Medium 1 containing 0.1% cholesterol, 2.0% glucose, 0.5% yeast extract, 0.2% NH₄NO₃, 0.02% K₂HPO₄ and 0.03% MgSO₄.7H₂O (pH 7.0) (Ahmad and Johri, 1992) was used. Bacterial cells were grown at 30°C with shaking at 150 rev min⁻¹.

Effect of different carbon sources for the production of cholesterol oxidase

Cholesterol oxidase production was investigated by modifying the medium with different carbon sources such as sucrose, fructose, maltose, glucose and xylose in the production medium at a concentration of 2.0%. The amount of enzyme produced by the *Bacillus cereus* strain KAVK4 at 12, 24, 32, 48, 64, 80hrs of incubation time was determined by the enzymatic assay.

Effect of different nitrogen sources for the production of cholesterol oxidase

The influence of different nitrogen sources on cholesterol oxidase production was investigated by substituting yeast extract, ammonium nitrate of basal medium with different organic and inorganic nitrogen sources such as beef extract, peptone, ammonium chloride, sodium nitrate and ammonium nitrate at a concentration of 0.5% and 0.2%. The amount of enzyme produced by the *Bacillus cereus* strain KAVK4 at 12, 24, 32, 48, 64, 80 hrs of incubation time was determined by the enzymatic assay.

Effect of different metal ions for the production of cholesterol oxidase

To investigate the effect of different metal ion on cholesterol oxidase production, cells were cultivated in the medium containing different metal ions such as zinc sulphate, manganese sulphate, ferrous sulphate, copper sulphate and magnesium sulphate at a concentration of 0.03%. The amount of enzyme produced by the *Bacillus cereus* strain KAVK4 at 12, 24, 32, 48, 64, 80 hrs of incubation time was determined by the enzymatic assay.

Effect of different pH for the production of cholesterol oxidase

In order to investigate the effect of pH on cholesterol oxidase production, the selected bacterial isolate was cultivated with different pH such as 6, 6.5, 7.0, 7.5, 8.0 in shake flask culture. The amount of enzyme produced by the *Bacillus cereus* strain KAVK4 at 12, 24, 32, 48, 64, 80 hrs of incubation time was determined by the enzymatic assay.

Effect of different temperature for the production of cholesterol oxidase

The influence of different temperature on cholesterol oxidase production was investigated using different temperature such as room temperature, 40°C, 50°C, 55°C, 65°C. The amount of enzyme produced by the *Bacillus cereus* strain KAVK4 at 12, 24, 32, 48, 64, 80 hrs of incubation time was determined by the enzymatic assay.

Effect of different time for the production of cholesterol oxidase

The optimized enriched media for the production of cholesterol oxidase was investigated using different time such as 12, 24, 32, 48, 64, 80 hrs of incubation time was determined by the enzymatic assay. Based on the effect of Carbon source, Nitrogen source, Metal ions source, pH, Temperature and Time; the media was selected for the maximum production of cholesterol oxidase.

RESULTS AND DISCUSSIONS

Bacillus cereus strain KAVK4 was isolated from butter and identified based on the morphological and biochemical characteristics (Table 1). *Bacillus cereus* was gram positive, which shows positive result on catalase and sugar fermentation test. Hence 16S rRNA gene region was amplified and sequenced to

identify and confirm the *Bacillus cereus* at molecular level and named the strain as KAVK4.

Table 1: Colony Morphology, Physical and Biochemical characterization of *Bacillus cereus*.

Colony Morphology	Cream, Smooth and Flat colonies
Physical Characterization	
Gram stain	Gram Positive
Biochemical characterization	
Catalase test	+
Oxidase test	-
Indole production	-
Citrate Test	+
Sugar fermentation	+

+ POSITIVE, - NEGATIVE

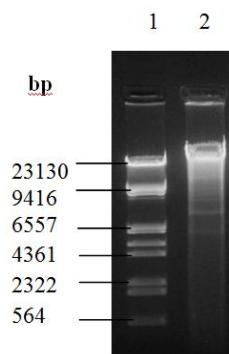


Fig 1: Genomic DNA of *Bacillus cereus*, 1- Lambda DNA / Hind III Digest Marker; 2- Genomic DNA of *Bacillus cereus* strain KAVK4.

Genomic DNA from *Bacillus cereus* strain KAVK4 was extracted (Figure 1). According to Jayachitra *et al.*, 2012, 16S rRNA gene region of *Bacillus cereus* was amplified and sequenced. 1.5 kb size of discrete bands was observed in 1% agarose gel on UV illuminator. PCR amplification of 16S rRNA gene region using Forward and Reverse primers, the obtained PCR product resulted in 1366 bp (Figure 2).

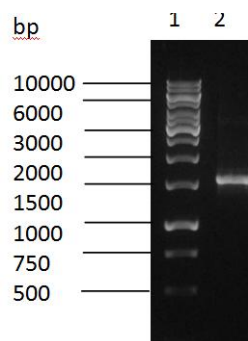


Fig 2: PCR Product of *Bacillus cereus*, 1- 1Kb DNA Ladder; 2-PCR product of *Bacillus cereus* strain KAVK4.

The sequence data was submitted to NCBI and the accession number is KP792775. This was assessed for cholesterol lowering potential. According to Bholay *et al.*, 2013, Cholesterol as sole source of carbon contained in the medium, the organism was inoculated in medium utilize the cholesterol and grown well. It must produce some enzymes which have ability to metabolite cholesterol. In my study nutrient agar medium with 1%

concentration of cholesterol was prepared and the isolates were inoculated. It utilize cholesterol and grown well on the medium (Figure 3).



Fig. 3: Cholesterol degradation of *Bacillus cereus* strain KAVK4.

The production of cholesterol oxidase is assessed by qualitative and quantitative method. Colony staining method and Cholesterol oxidase indicator plate are used for qualitative method of analysis. This was confirmed with the development of red color (Figure 4).



Fig. 4: Colony staining method of *Bacillus cereus* strain KAVK4.

In the indicator plate method, the medium color turns into brown because of the formation of azo component that was due to the presence of H_2O_2 generated by cholesterol oxidase (Figure 5) (Lashkarian *et al.*, 2010).



Fig. 5: Cholesterol oxidase indicator plate of *Bacillus cereus* strain KAVK4.

According to Kim *et al.*, 2002, *Bacillus subtilis* from Korean traditional fermented flatfish, produced cholesterol oxidase in the medium containing 0.2% cholesterol. In fermented flatfish,

the residual cholesterol was reduced by degrading potential of *Bacillus subtilis* due to the cholesterol oxidase production in the presence of oxygen (Kim *et al.*, 2003). Based on this concordance, *Bacillus cereus* strain KAVK4 was confirmed for degradation of cholesterol. The optimization of Medium is required to improve cholesterol oxidase production. Based on these the carbon source, Organic and inorganic nitrogen source, metal ion source, pH, temperature and time were modified for the maximum production of cholesterol oxidase. Some microorganisms have ability to grow and produce cholesterol oxidase in the presence of Tween 80. Some microorganisms were effectively produced cholesterol oxidase based on the temperature. The optimum temperature for this enzyme production *Rhodococcus equi* was at 47°C and *Corynebacterium cholesterolicum* production of this enzyme was at 40°C respectively (Salva *et al.*, 1999). *Streptomyces fradiae* and *Brevibacterium sp.* showed optimum stability for 30min at 50°C and 53°C respectively (Yazdi *et al.*, 2001b; Fujishiro *et al.*, 2002). The optimum pH for the enzyme activity was between 7.0 and 8.0 was observed in *Actinomyces lavendulae* mycelium, *Corynebacterium cholesterolicum*, *Streptovercillium cholesterolicum*, *Rhodococcus equi* and *Streptomyces violascens* (Lashkarian *et al.*, 2010). Among the five different media, media 1 showed maximum production was about 0.75 U/ml and was selected for further modification to enhance the production of cholesterol oxidase (Figure 6).

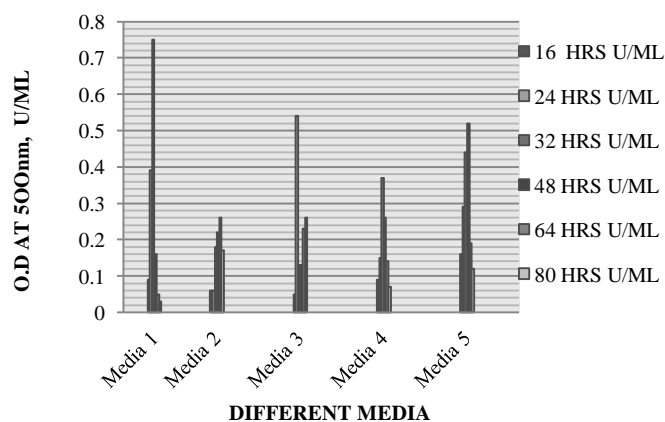


Fig. 6: Effect of different media for the production of cholesterol oxidase.

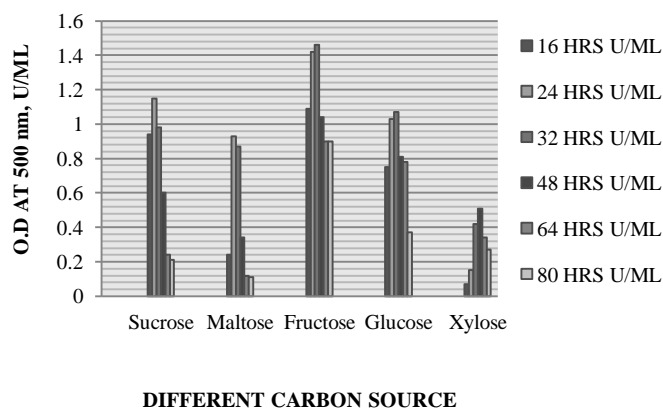


Fig. 7: Effect of different carbon source for the production of cholesterol oxidase.

Fructose enriched media showed highest production was about 1.46 U/ml (Figure 7). In the organic and inorganic nitrogen source of media, Beef extract and Ammonium nitrate enriched media were produced cholesterol oxidase was about 1.05 and 1.42 U/ml respectively (Figure 8).

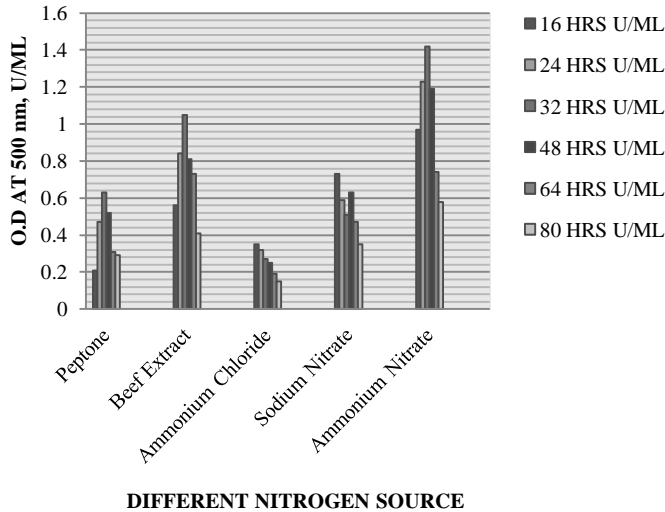


Fig. 8: Effect of different nitrogen source for the production of cholesterol oxidase.

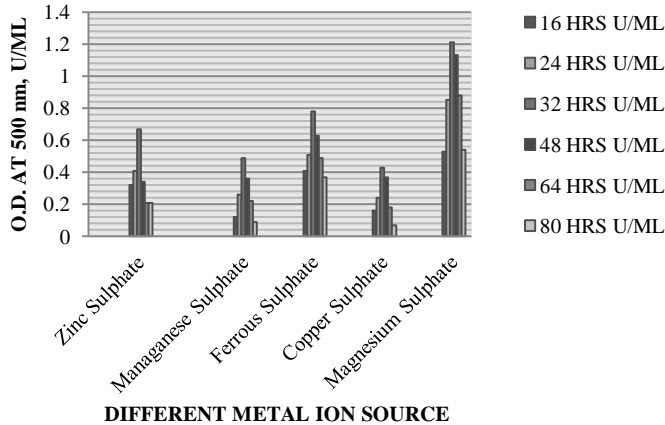


Fig. 9: Effect of different metal ion source for the production of cholesterol oxidase.

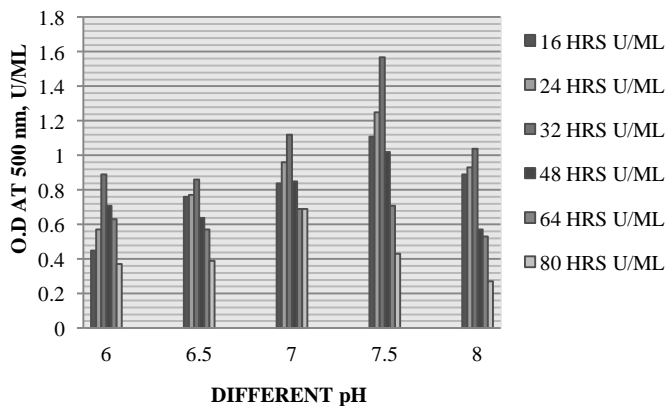


Fig. 10: Effect of different pH for the production of cholesterol oxidase.

For the metal ion source, magnesium sulphate showed maximum of about 1.21 U/ml respectively (Figure 9). The optimum pH for the production of cholesterol oxidase was at pH 7.5 of 1.57 U/ml (Figure 10) and the temperature required for the optimum production of cholesterol oxidase at Room temperature was about 0.97 U/ml (Figure 11).

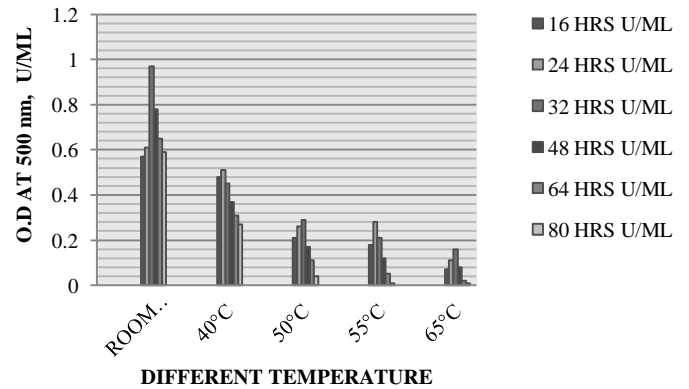


Fig. 11: Effect of different temperature for the production of cholesterol oxidase.

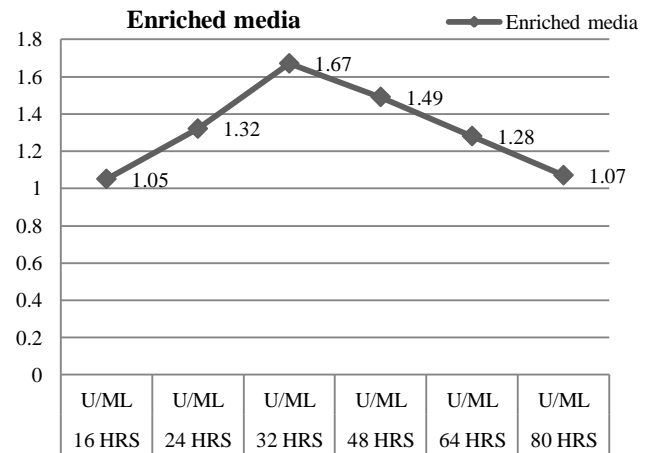


Fig. 12: Effect of different time for the production of cholesterol oxidase.

The enriched medium which showed maximum production was selected for further study. Hence the *Bacillus cereus* strain KAVK4 showed maximum production of 1.67 U/ml at 32 hours of incubation period (Figure 12).

CONCLUSION

Application of cholesterol oxidase is commercial and medical important. For this reason, the *Bacillus cereus* strain KAVK4 which are capable of producing maximum levels of cholesterol oxidase have been isolated and identified. For better identification, 16S rRNA gene region was amplified and sequenced to confirm the species at molecular level. The optimization of cholesterol oxidase production by *Bacillus cereus* strain KAVK4 should prove that increase the oxidation of

cholesterol and converts into 4-cholesten-3-one and other derivatives with the reduction of O₂ to H₂O faster.

REFERENCES

- Ahmad S, Johri BN. Sterol bioconversion using *Rhodococcus equi* in organic-aqueous two-lipid phase system. Indian J Microbiol, 1992; 32: 489–491.
- Allain CC, Poon LS, Chan CS, Richmond W, Fu PC. Enzymatic determination of total serum cholesterol. Clin Chem, 1974; 20(4):470–475.
- Arima K, Nagasawa M, Bae M, Tamura G. Microbial transformation of steroids. Part I. Decomposition of cholesterol by microorganisms. Agric Biol Chem. 1969; 33:1636-1643.
- Bholay AD, Gadekar DJ, Sawant SK, Sonawane SM. Bacterial extracellular cholesterol oxidase and its pharmaceutical perspectives. Int J Curr Microbiol App Sci, 2013; 2(3): 19-28
- Constantinides A. Steroid transformation at high substrate concentrations using immobilized *Corynebacterium simplex* cells. Biotechnol Bioeng, 1980; 22(1): 119-136.
- Fujishiro K, Uchida H, Shimokawa K, Nakano M, Sano F, Ohta T, Nakahara N, Aisaka K, Uwajima T. Purification and properties of a new *Brevibacterium sterolicum* cholesterol oxidase produced by E.coli MM294/pnH10. J FEMS Microbiol Lett, 2002; 215 (2): 243–248.
- Jayachitra A, Krithiga N, Bavani C. Isolation, molecular characterisation and sequencing of cholesterol degrading bacteria. Int J Pharm Life Sci, 2012; 3(9): 1946-1952
- Kaunitz H. Degradation of cholesterol oxidase by *Bacillus subtilis* SFF34 in flatfish during fermentation. Lipids, 1978; 13: 373-375.
- Kim KP, Rhee CH, Park HD. Degradation of cholesterol by *Bacillus subtilis* SFF 34 isolated from Korean traditional fermented flatfish. Lett Appl Microbiol, 2002; 35: 468-472.
- Kim KP, Rhee IK, Park HD. Degradation of Cholesterol by *Bacillus Subtilis* SFF 34 in flatfish during fermentation. J Microbiol, 2003;41(4): 284-288.
- Lashkarian H, Raheb J, Shahzamani K, Shahbani H, Shamsara M. Extracellular cholesterol oxidase from *Rhodococcus* sp: Isolation and molecular characterization. Iran Biomed J, 2010; 14 (1-2): 49-57.
- Lata Kumari, Shamsher S. Kanwar. Cholesterol oxidase and its applications. Adv Microbiol, 2012; 2: 49-65
- Ming-Tsung Lee, Wen-Chang Chen, Cheng-Chun Chou. Medium improvement by orthogonal array designs for cholesterol oxidase production by *Rhodococcus equi* No. 23. Process Biochem, 1997; 32(8): 697-703
- Pitcher DG, Saunders NA, Owen RJ. Rapid extraction of bacterial genome DNA with guanidium thiocyanate. Lett Appl Microbiol, 2008; 8(4):151-156.
- Sakodinskaya IK, Alexander D. Ryabov. Crown ether activates cholesterol oxidase in low water media. Biotechnol Lett, 2000; 22: 173-176.
- Salva TJG, Liserre AM, Moretto AL, Zullo MAT, Ventrucci G, Menezes TJB. Some enzymatic properties of cholesterol oxidase produced by *Brevibacterium* sp. Revista de Microbiologia, 1999; 30 (4): 315-323.
- Smith AG, Brooks CJW. Cholesterol oxidase: properties and application. J Steroid Biochem, 1976; 7: 705-713.
- Varma R, Nene S. Biosynthesis of cholesterol oxidase by *Streptomyces lavendulae* NCIM 2421. Enzyme microb Technol, 2003; 33: 286-291.
- Watanabe K, Shimizu H, Aihara H, Nakamura R, Suzuki KI, Momagata K. Isolation and identification of cholesterol degradation *Rhodococcus* strains food of animal origin and their cholesterol oxidase activities. J Gen Appl Microbiol, 1986; 32: 137-147.
- Williams and Wilkins., 9th Edition. Bergey's Manual of determinative bacteriology, 1994
- Wu K, Li W, Song J, Li T. Production, purification, and identification of Cholest-4-en-3-one produced by Cholesterol oxidase from *Rhodococcus* sp. in aqueous/organic biphasic system. Biochem Insights, 2015; 8(S1): 1–8 doi:10.4137/BCI.S21580.
- Yazdi MT, Yazdi ZT, Ghasemian A, Zarrini G, Olyaei NH, Sephehrizadeh Z. Purification and characterization of extra-cellular cholesterol oxidase from *Rhodococcus* sp. PTC1633. Biotechnology, 2008; 7(4):751–756.
- Yazdi MT, Malekzadeh F, Zarrini GH, Faramarzi MA, Kamranpour N, Khaleghparast SH. Production of cholesterol oxidase by a newly isolated *Rhodococcus* sp. World J Microbiol Biotechnol, 2001a; 17: 731-737.
- Yazdi MT, Zahraei M, Aghaeipour K, Kamranpour N. Purification and partial characterization of cholesterol oxidase from *Streptomyces fradiae*, Enzyme Microb Tech, 2001b; 18 (4-5):410-414.

How to cite this article:

Amutha K, Kokila V. Optimization of cholesterol oxidase production and 16S rRNA partial sequence of *Bacillus cereus* strain KAVK4 isolated from butter. J App Pharm Sci, 2016; 6 (07): 061-066.