**Antifungal activity of *Terminalia chebula* extract against wilt fungi *Fusariumoxysporum*an important plant pathogen**

**Padmalatha A, Dr. Prabha S.B**

Department of Microbiology, Vel’s Institute of Science, Technology and Advanced Studies (VISTAS),

PV Vaithiyalingam Rd, Velan Nagar, Krishnapuram, Pallavaram, Chennai 600117, Tamil Nadu. India.

\*Corresponding Author E-mail: **apadmalatha@gmail.com**

**ABSTRACT:**

*Terminalia chebula,* a deciduous plant, endogenous to India has medicinal value. Its fruits are known for its cure to many ailments in human beings. This study is to test the Biological activity of these compounds tested against plant pathogenic fungai *Fusarium oxysporum*which cause wilt in economically important crops. Wilt is a devastating disease whichdestroys the infected plant and cause economic loss to farmers. Panama disease in bananas is a type of wilt disease which is very prevalent in India. This fungai has broad host range from tomato, cotton etc. Outbreak reported in many parts of India. Standard culture of *Fusarium oxysporum* ITCC -6246, *Fusarium oxysporum f. sp. ciceri* ITCC 3636 and *Fusarium oxysporum f. sp. Lycopersici* MTCC 10270 were used to detect the potential chemical compound in dried fruitby macro-dilution.Each organic solvent (Water, Ethanol, Chloroform, Benzene) has different extraction capacity based on the polarity of chemical compound in plants. These chemical compounds extensively studied for their antioxidant property and preliminary screening of chemical compound constituents in all extract. Mycelium growth inhibition tested on solid medium and inhibition measured in mm diameter. Spores of *Fusarium*sp. present in soil for many months in dormant form. Under suitable climatic conditions this will germinate and cause infection in plants.  Rate of spore germination inhibition in liquid medium measured and calculated for every solvent extract. Rate of inhibition was calculated and analysed statistically by t test. The efficient solvent extract determined for bulk extraction of biologically active compound from seeds.

**KEYWORDS:** Phytochemical, fungicides, plant pathogen, macro-dilution, plant diseases.

**INTRODUCTION:**

Phytochemical are plant based substances produced by the plant for the plant to protect itself from the adverse condition and from impact of mircoorganisms, animals, insects etc1,2,3. They are considered non-nutritive but biologically active compound. Phytochemicals are well known for its beneficial4,5,6,7,8effect but some are destructive. They are distinct to each plant even specific to its part. Varied climatic condition in India influence the production of secondary phytochemical in plants. Phytochemical are not only used as medicine but also used as nutritional supplements, enhance taste, flavour and colour.

Phytochemicals based on their chemical nature are divided in to many categories, carbohydrates, lipids, phenolic compounds, terpenoids and alkaloids9,10,11.  Each of these compound has varying degree of antioxidant property12, biological activity and it is remedy for many ailments. Pharmaceutical products like suspension prepared with plant based antioxidant to improve self-life.

Phenolic compound contain benzene rings withsimple hydroxyl groupto highly complex side chain. These secondary metabolite broadly classified under flavonoids, phenolic acids, xanthons, lignans and anthocyanins. Phenolic compounds helps to pollinate, protect from animals and antimicrobial effect13,14,15,16. Terpenoids otherwise known as isoprenoids impart flavour and fragrance known to have antimicrobial activity, ithelps the plant to interact with environment and tolerance to adverse condition eg. Vitamin A. Alkaloids natural base contain nitrogen, reserve nitrogen acts as growth stimulator and mainly involved in plant defence. They are soluble in water under acidic, in neutral or basic condition soluble in lipids17. These molecules have wide range of application18.

In this study inhibitory activity of extracted compound from seed of *Terminalia chebula* tested for spore germination of plant pathogenic fungai, *Fusarium oxysporum*(Wilt in Tomato, Banana, Guava), *Fusarium oxysporum*f. sp. ciceri (Wilt in Chickpea) and *Fusarium oxysporum* f. sp. Lycopersici (Wilt in tomato).

Management of these plant diseases is a complexactivity as there is always need of new compound to control the growth19 of fungi targeting either mycelium or spore germination without affecting the ecosystem20. Many scientist are interested in phytochemical to control plant disease, tested many compounds against plant pathogenic microorganisms by in vitro found capable of controlling the growth21.

**MATERIALS AND METHODS:**

**Sample collection and processing:**

Terminalia chebula dried fruit sample were collected from Chennai, Tamil Nadu. Surface washed with sterile distilled water and air dried. These fruits were broken and divided into two parts, only seeds collected and powdered. Hundred grams of powdered *Terminalia chebula* seed materials were weighed into separate sterile conical flasks. The samples were extracted using 500 ml of aqueous, ethanol, chloroform and benzene separately and left for 48 hours at room temperature. The resultant suspensions were filtered into sterile conical flasks. These extracts were air dried and the residue was collected15if needed suspend in sterile double distilled water.

**Preliminary phytochemical analysis:**

Phytochemical composition22, 23 of sample tested for sugars, amino acids, anthroquinins, coumarins, saponins, tannins, phenolic compounds, flavonoid, terpenoids and glycosides24results were tabulated (Table 1).

**DPPH (2, 2-diphenyl-1-picrylhydrazyl) radical scavenging assay:**

DPPH radical scavenging assaywas performed to test presence of antioxidants4,13,14,16. In brief, 0.135 mM DPPH was prepared in methanol. Different concentration of extract (5, 10, 20, 40, 80, 160 and 320 μg/ml) was mixed with 2.5 ml of DPPH solution. The reaction mixture was vortexes thoroughly and kept at room temperature for 30 min. Ascorbic acid was used as the reference standard. The Absorbance of the mixture was measured at 517 nm. The ability of plant extract to scavenge DPPH radical and control was calculated from the following formula: respectively.

Rate of DPPH inhibition = [(absorbance of control - absorbance of test)/(absorbance of control)] ×100

**Preliminary study for antifungal activity by well diffusion:**

Seven day old culture of Fusariumoxysporum seeded by spread-plate method on Mueller-Hinton agar. Approximately wells of uniform size (0.65 cm) were made with a cork-borer onto the plates inoculated with test organisms. Crude plant extracts of 250 μl, 500 μl, 500μl and 1000μl were respectively added into each well aseptically. Triplicates were used as replicates for each treatment. Inoculated plates were incubated at room temperature until the fungal growth of the control plates reached the edge of the plate. Zone of inhibition measured and mean calculated for further analysis.

**Antifungal activity by dilution method:**

Standard fungal plant pathogen were selected to test the ability of seed extract for their potential antifungal capabilities. *Fusarium oxysporum* ITCC -6246, *Fusarium oxysporum* f. sp. ciceri ITCC 3636 and *Fusarium oxysporum* f. sp. lycopersici MTCC 10270 was obtained from the Microbial Type Culture Collection (MTCC), Chandigarh, India and Indian type culture collection (ITCC), New Delhi, India. Maintained the strains on Potato dextrose agar at 4°C.Fifty ml of sterile distilled water was added to 7 day old culture grown on potato dextrose agar (PDA) and scraped gently, filtered through Buchner funnel under sterile condition. The filtrate was adjusted to 1x105 spores/ml and used for further study. 100µl of conidial suspension, 100µl of *Terminalia chebula* (ethanol, chloroform, benzene, water) extracts of seed with various concentrations ranging from 1.0, 2.0, 5.0 mg/mL were added separately and incubated in a shaker at 28°C with 120 rpm for 10 days. For control, test tube received 100µl of conidial suspension and 100µl of sterile distilled water. Optical readings were taken at 600 nm at every 2days interval up to 10 days. All procedures described above were carried out under sterile conditions. Experiments were done in triplicate mean was calculated, rate of inhibition was calculated by (aborbance of control-abosrbance of test sample)/ (aborbance of control \*100).

**RESULTS AND DISCUSSION:**

**1.     Phytochemical constituents of extracts:**

Various chemical constituents expressed as high, medium, low and nil for four solvent mediums.

**2.     Antioxidant assay:**

Ethanol extract shows maximum and benzene extract shows least antioxidant activity.Polar compounds from the dried fruit exhibit higher antioxidant activity than nonpolar compound extracted with benzene (Fig.1)

**Table 1: Phytochemical analysis of *Terminalia chebula* seeds for the following constitutions**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Plant constitutions** | **Seed extract with solvent** | | | |
| **Aqueous** | **Ethanol** | **Chloro**  **form** | **Benzene** |
| Steroids | + | +++ | ++ | - |
| Terpenoids | +++ | - | ++ | + |
| Reducing sugar | - | +++ | ++ | + |
| Sugars | - | - | - | ++ |
| Phenolic compound | ++ | +++ | + | - |
| Flavonoids | - | - | - | - |
| Saponins | - | - | ++ | +++ |
| Tannins | - | + | +++ | +++ |
| Amino acids | + | +++ | ++ | - |
| Glycoside | - | +++ | - | ++ |
| Coumarins | - | +++ | +++ | + |
| Anthroquinones | +++ | ++ | - | - |

**Fig 1: DPPH method of antioxidant assay of *Terminalia chebula* dried seed extract.**

**3.     Well diffusion:**

Mean value of diameter of Zone of inhibition in mm of different solvent extract on *Fusarium oxysporum*ITTC 6246 in table 2.

**Table 2 Zone of inhibition in mm against *Fusarium oxysporum* ITCC 6246 of different solvent extract**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Solvent extract | 250 μl | 500 μl | 750 μl | 1000 μl |
| Water | 0 | 0 | 0 | 0 |
| Ethanol | 13.06 | 14.63 | 14.76 | 21.03 |
| Chloroform | 12.06 | 13.2 | 15.06 | 18.1 |
| Benzene | 0.53 | 0.66 | 21.13 | 28.06 |

**Fig 2: Zone of inhibition of various Organic solvent extract water, ethanol, chloroform and benzene against *Fusarium oxysporum* ITCC6246**

**4.     Rate of inhibition:**

Spore germination inhibition of *Fusarium oxysporum* ITCC -6246, *Fusarium oxysporum f. sp. ciceri* ITCC 3636 *and Fusarium oxysporum f. sp. lycopersici* MTCC 10270 absorbance measured in UV spectrophotometer, from mean value rate of inhibition tabulated (Table 2, 3 and 4) rate of inhibitory concentration (IC) range from 17 to 93. IC 70 considered as most efficient.

**Table 3: Rate of inhibitory concentration of *Terminalia chebula* seed extract against *Fusarium oxysporum* ITCC -6246 using different solvent**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Solvent | Days | Negative |  |  |  |
|  | Control | 1.0 mg/mL | 2.0 mg/mL | 5.0 mg/mL |
|  | 2 | 0.264 | 47.34 | 64.77 | 70.07 |
| Water | 4 | 0.408 | 40.44 | 51.71 | 62.5 |
|  | 6 | 0.671 | 48.88 | 54.39 | 64.08 |
|  | 8 | 0.898 | 36.63 | 41.75 | 51.33 |
|  | 10 | 0.989 | 19.21 | 28.87 | 41.45 |
|  | 2 | 0.264 | 66.66 | 80.68 | 91.28 |
| Ethanol | 4 | 0.408 | 66.17 | 75.98 | 85.04 |
|  | 6 | 0.671 | 70.78 | 77.94 | 86.14 |
|  | 8 | 0.898 | 69.59 | 77.83 | 84.74 |
|  | 10 | 0.989 | 66.53 | 72.29 | 81.69 |
|  | 2 | 0.264 | 59.09 | 74.24 | 85.6 |
| Chloroform | 4 | 0.408 | 57.35 | 63.72 | 77.69 |
|  | 6 | 0.671 | 59.16 | 63.18 | 73.47 |
|  | 8 | 0.898 | 47.88 | 57.34 | 65.36 |
|  | 10 | 0.989 | 37.61 | 49.14 | 51.66 |
|  | 2 | 0.264 | 57.95 | 66.28 | 74.62 |
| Benzene | 4 | 0.408 | 46.81 | 57.59 | 67.4 |
|  | 6 | 0.671 | 52.45 | 58.56 | 65.57 |
|  | 8 | 0.898 | 40.86 | 48.99 | 55.56 |
|  | 10 | 0.989 | 25.37 | 35.08 | 43.07 |

**Table 4: Rate of inhibitory concentration of Terminalia chebula seed extract against *Fusarium oxysporum* f. sp. ciceri ITCC 3636 using different solvent**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Solvent** | **Days** | **Negative** |  |  |  |
|  | **Control** | **1.0 mg/mL** | **2.0 mg/mL** | **5.0 mg/mL** |
|  | 2 | 0.243 | 45.67 | 55.96 | 60.08 |
| Water | 4 | 0.314 | 25.47 | 35.35 | 53.18 |
|  | 6 | 0.597 | 40.87 | 50.08 | 51.25 |
|  | 8 | 0.863 | 33.83 | 42.52 | 53.88 |
|  | 10 | 0.971 | 17.71 | 21.93 | 38.61 |
|  | 2 | 0.243 | 76.13 | 80.24 | 93 |
| Ethanol | 4 | 0.314 | 51.27 | 58.28 | 71.65 |
|  | 6 | 0.597 | 60.3 | 70.18 | 77.05 |
|  | 8 | 0.863 | 63.26 | 72.88 | 80.99 |
|  | 10 | 0.971 | 61.99 | 71.36 | 79.6 |
|  | 2 | 0.243 | 60.08 | 74.07 | 79.01 |
| Chloroform | 4 | 0.314 | 37.26 | 45.54 | 58.91 |
|  | 6 | 0.597 | 49.91 | 58.29 | 60.13 |
|  | 8 | 0.863 | 45.19 | 51.68 | 59.32 |
|  | 10 | 0.971 | 34.7 | 44.59 | 59.93 |
|  | 2 | 0.243 | 51.02 | 63.37 | 69.65 |
| Benzene | 4 | 0.314 | 29.61 | 42.03 | 57.96 |
|  | 6 | 0.597 | 44.89 | 54.27 | 62.98 |
|  | 8 | 0.863 | 37.77 | 43.91 | 56.89 |
|  | 10 | 0.971 | 25.12 | 35.42 | 47.16 |

**Table 5: Rate of inhibitory concentration of *Terminalia chebula* seed extract against *Fusarium oxysporum* f. sp. lycopersici MTCC 10270 using different solvent**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Solvent** | **Days** | **Negative** |  |  |  |
|  | **Control** | **1.0 mg/mL** | **2.0 mg/mL** | **5.0 mg/mL** |
|  | 2 | 0.239 | 50.62 | 63.59 | 71.96 |
| Water | 4 | 0.304 | 25 | 44.73 | 58.22 |
|  | 6 | 0.447 | 27.29 | 37.13 | 50.11 |
|  | 8 | 0.743 | 27.72 | 32.57 | 44.68 |
|  | 10 | 0.963 | 18.06 | 27.62 | 41.95 |
|  | 2 | 0.239 | 78.66 | 86.19 | 89.95 |
| Ethanol | 4 | 0.304 | 68.09 | 77.96 | 86.51 |
|  | 6 | 0.447 | 70.02 | 82.55 | 86.12 |
|  | 8 | 0.743 | 74.83 | 82.36 | 88.02 |
|  | 10 | 0.963 | 65.31 | 77.46 | 88.36 |
|  | 2 | 0.239 | 71.54 | 78.66 | 87.44 |
| Chloroform | 4 | 0.304 | 86.5 | 55.92 | 61.51 |
|  | 6 | 0.447 | 45.63 | 50.55 | 61.74 |
|  | 8 | 0.743 | 57.33 | 60.02 | 63.52 |
|  | 10 | 0.963 | 46.62 | 51.92 | 62.82 |
|  | 2 | 0.239 | 58.99 | 71.96 | 77.4 |
| Benzene | 4 | 0.304 | 37.82 | 50.32 | 61.18 |
|  | 6 | 0.447 | 41.61 | 40.26 | 53.46 |
|  | 8 | 0.743 | 40.64 | 38.49 | 47.91 |
|  | 10 | 0.963 | 24.09 | 34.78 | 49.53 |

5. **Efficacy of crude seed extract.**

Around seventy percent of spore germination inhibition rate considered as significant activity of seed extract (mg/ml) against plant pathogenic fungi. Initial incubation with ethanol extract effectively inhibit *Fusarium oxysporum*ITCC -6246, *Fusarium oxysporum f. sp. ciceri*ITCC 3636 and *Fusarium oxysporum f. sp. lycopersici*MTCC 10270spore germination. 1mg/ml Effective minimum inhibitory concentration of ethanol extract against *Fusarium oxysporum f. sp. Lycopersici*MTCC 10270, t test performed in excel p value less than 0.05 confirms the significant antifungal activity.

**CONCLUSION:**

Terminalia chebula seed extract possess antifungal activity against *Fusarium oxysporum sp.*Based on these analysis ethanol extractalternate for synthetic compound controlling plant fungal disease. Furtherchloroform extract contain biologically active phytochemical which will play major contribution in controllingmycelium growth. Phytochemical found in these extract used to control wilt fungi spore germination as well as mycelial growth repectively.

**CONFLICT OF INTEREST:**

The author(s) declare no conflict of interest.

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**REFERENCES:**

1.      Bode A.M. and Dong Z. Toxic phytochemicals and their potential risks for human cancer. Cancer Prevention Research. 2015; 8(1): https://doi.org/10.1158/1940-6207.capr-14-0160

2.      ErmiAbriyani. Lia Fikayuniar. Screening Phytochemical, Antioxidant Activity and Vitamin C Assay from Bungoperak-perak (Begonia versicolarIrmsch) leaves. Asian J. Pharm. Res. 2020; 10(3): https://doi.org/10.5958/2231-5691.2020.00032.5

3.      Gul R. Jan S.U. Faridullah S. Sherani, S. and Jahan N. Preliminary phytochemical screening, quantitative analysis of alkaloids, and antioxidant activity of crude plant extracts from Ephedra intermedia indigenous to Balochistan. The Scientific World Journal, 2017; https://doi.org/10.1155/2017/5873648

4.      Martinez K.B. Mackert J.D. and McIntosh M.K. Polyphenols and intestinal health. In Nutrition and Functional Foods for Healthy Aging. 2017; 191-210. Academic Press. https://dx.doi.org/10.1016/j.jnutbio.2015.12.021

5.      Sabarinath C, Sudhakar P, Shanmuganath C. Phytochemical and Antibacterial screening on leaves of Solanumtorvum. Asian J. Res. Pharm. Sci. 2018; 8(3):130-132.

6.      SarmisthaRej. Madhurima Dutta. Shahid Jamal. Sumanta Das. Sabyasachi Chatterjee. Study of Phytochemical Constituents and Antibacterial Activity of Clerodendruminfortunatum. Asian J. Res. Pharm. Sci. 2014; 187-195.

7.      Singh M.K. Singh S.K. Singh AV. Verma H. Singh P.P. and Kumar A. Phytochemicals: Intellectual property rights. In Functional and Preservative Properties of Phytochemicals. 2020; 363-375. Academic Press.

8.      Suresh G. Killedar. Harinath N. More. Screening of Antimicrobial Potential and Phytoconsituents for Different Extracts of MemecylonumbellatumBurm Inflorescences. Asian J. Pharm. Res. 2011; 114-118.

9.      Batra Neha. Jain Honey. BairwaRanjan. BachwaniMukesh. Pharmacognostical and preliminary phytochemical investigation of Acoruscalamuslinn. Asian J. Pharm. Res. 2012; 39-42.

10.    Huang Y. Xiao D. Burton-Freeman B.M. and Edirisinghe I. Chemical changes of bioactive phytochemicals during thermal processing. 2016: https://doi.org/10.1016/B978-0-08-100596-5.03055-9

11.    Mohanty PK. Neha Chourasia. Neraj Kumar Bhatt YA. Jaliwala. Preliminary Phytochemical Screening of Cajanuscajan Linn. Asian J. Pharm. Tech. 2011; 49-52.

12.    Valli G. Jeyalakshmi M. Preliminary Phytochemical and Antioxidant Study of Odina woodier Leaf Extract. Asian J. Pharm. Res. 2012; 153-155.

13.    Lin D. Xiao M. Zhao J. Li Z. Xing B. Li X. Kong M. Li L. Zhang Q. Liu Y. and Chen H. An overview of plant phenolic compounds and their importance in human nutrition and management of type 2 diabetes. Molecules. 2016: 21(10). https://doi.org/10.3390/molecules21101374

14.    Perumal P.A.R.T.H.A.S.A.R.A.T.H.I. and Saravanabhavan, K.A.V.I.T.H.A. Antidiabetic and antioxidant activities of ethanolic extract of Piper betle L. leaves in catfish, Clariasgariepinus. Asian J Pharm Clin Res. 2018; 11(3). http://dx.doi.org/10.22159/ajpcr.2018.v11i3.22393

15.    Umeh EU. Oluma HOA and Igoli JO. Antibacterial screening of four local plants using an indicator-based microdilution technique. African Journal of Traditional, Complementary and Alternative Medicines. 2005; 2(3). https://doi.org/10.4314/ajtcam.v2i3.31121

16.    Vuolo MM. Lima VS. and Junior MRM. Phenolic compounds: Structure, classification, and antioxidant power. In Bioactive Compounds. 2019; 33-50. Woodhead Publishing. https://doi.org/10.1016/B978-0-12-814774-0.00002-5

17.    Verpoorte R. 2005. Alkaloids.

18.    Jaiganesh KP. Sreedharren B. ArunachalamG. Nirmala R. Nepolean R. Pharmacognostical and Antimicrobial investigation of Jatropha curcas, Linn., Leaf. Asian J. Res. Pharm. Sci. 2013; 3(4).

5.      Rahman M. and Borah M. Phytochemicals: Their role and mechanism in suppressing plant pathogenic bacteria. 2021; https://doi.org/10.22271/tpi.2021.v10.i6g.6560

19.    Padmalatha A. Control of plant disease by biological agent. International Journal of Multidisciplinary Educational Research. 2021; 6(9).  http://ijmer.in.doi./2021/10.06.167

20.    Barbieri R. Coppo E. Marchese A. Daglia M. Sobarzo-Sánchez E. Nabavi S.F. and Nabavi S.M. Phytochemicals for human disease. An update on plant-derived compounds antibacterial activity. Microbiological research. 2017; 44-68.   https://doi.org/10.1016/j.micres.2016.12.003

21.    Rakshit Pathak. Anjali Kumari. Mohammed Mohsin. Ganga Bisht. MadhuBala. Phytochemical Assessment and In vitro Antioxidant potential of Camelinasativa L. seed cake; Asian J. Research Chem. 2020; 13(1). https://doi.org/10.5958/0974-4150.2020.00009.7

22.    RangarajanNarasimhan, Sathiyamoorthy. M. Phytochemical Screening and Antioxidant Studies in the Pulp Extracts of Cucurbita maxima. Asian J. Pharm. Res.  2016; 6(1): https://doi.org/10.52711/0974-360X.2021.00356

23.    Jamal Basha D. Srinivas Murthy BR. Prakash P. Anuradha KC. Preliminary Phytochemical Investigation and In Vitro Anti Bacterial Studies of Polycarpaeaaurea Wight and Arn (Caryophyllaceae) Emulgel. Asian J. Pharm. Res. 2018; 8(3): https://doi.org/10.5958/2231-5691.2018.00027.8

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